



#### Journal of Global Pharma Technology

Available Online at: www.jgpt.co.in

**RESEARCH ARTICLE** 

# Staphylococcus aureus Enterotoxin as a Major Risk Factor for Multiple Sclerosis Severity

# Suhaib Khalid Ibrahim<sup>1\*</sup>, Thanaa Rasheed Abdulrahman<sup>2</sup>, Nawfal Madhi Sheaheed<sup>3</sup>

- <sup>1</sup> MSc Microbiology / College of Health and Medical Technology / Baghdad / Iraq.
- <sup>2</sup> PhD. Microbiology, Al-Nahrain University / College of Medicine / Iraq.
- 3. Consultant Neurologist, Baghdad Teaching Hospital, Medical City/Iraq.

#### \*Corresponding Author: Suhaib Khalid Ibrahim

#### Abstract

Background: Staphylococcus aureus produce enterotoxins that function as super antigens which activate auto reactive CD4+ T-cells potentially target the basic myelin protein into the CNS. Objective: This study investigates the correlation between the colonization of S. aureus harbouring enterotoxins with multiple sclerosis (MS) exacerbation. Methods: A total of 200 nasal swabs were collected from three study groups: Healthy controls as a Non-MS Subjects (n=100), relapsing remitting MS or exacerbated (n=50) and newly diagnosed MS. (n=50). S. aureus was isolated from the anterior nares of these groups following standard operating procedures. Antibiotic susceptibility test against 19 antibiotics using disk diffusion method were done. Staphylococcal super Antigen SeA, seB, seC genes were amplified using standard conventional polymerase chain reaction (PCR) technique and resolved by agarose gel electrophoresis. Results: In this study, Out of 100 MS patients and 100 healthy control groups, there were 36% and 46% males and 64% and 54% females respectively. A total of 81(81%) were colonized with S. aureus includes, 31(38.2%) newly diagnosed MS and 50(61.8%) exacerbated MS while only 12(12%) isolated from control non-MS group. All S. aureus isolates were resistant to Methicillin (100%) and sensitive to Impenem (100%). PCR results showed that, the frequency of enterotoxins (sea, seb and sec) in MS patients was (42.4%) including (40.8%) were from newly diagnosed and (43.3%) were from relapsing remitting while there is no enterotoxins was detected in control group, the prevalence of se A was significantly higher (p<0.00) in the MS exacerbation (72%) than in newly diagnosed group (64.5%). Conclusions: The frequency of S. aureus isolates and its enterotoxin seA gene is high in the MS patients and this gene serve as an important marker in the severity of MS disease. Also S. aureus isolates were sensitive to imipenem which is considered as a better choice for nasal decolonization of S. aureus in the MS patients.

Keywords: Multiple sclerosis (MS), Staphylococcus aureus, Super antigens, Enterotoxin A, B, C.

#### Introduction

Multiple sclerosis (MS) is an autoimmuneinflammatory disease that occurs in the central nervous system (CNS) causing demyelination. The demyelination operation is related to infiltrating T-cells particular for major myelin proteins of the CNS, this disease has a peak start between 20 and 40 years old and influences women almost two times as often as men[1]. The most common form of disease is the relapsing-remitting MS (RR-MS), including around 85 - 90% of all cases. Although the etiology of MS is unknown; genetic and environmental factors play a role. Infectious pathogens are the possible environmental factors contributed to

the MSdevelopment orseverity [2].Pathogens related to the severity development of MS involve bacteria; such as Chlamydia pneumoniae Mycoplasmapneumonia and Staphylococcus aureus which produced enterotoxins that function as superantigens [3]. It has been proposed that S. aureus in nasal carriage is related to several autoimmune diseases systemic lupus erythematosus, rheumatoid and Wegener's granulomatosis arthritis syndrome through superantigens such as toxic shock syndrome toxin-1 {tsst-1} or staphylococcal enterotoxins (SEs) [4].

Superantigens are proteins produced mostly by bacteria and some viruses that potently activate autoreactive CD4+ T cells, inducing massive cell proliferation and cytokine production. predominantly IL-2interferon (IFN)-y [5]. Staphylococcus aureus superantigens are capable of activating T cells due to their ability to bind to both major histocompatibility complex (MHC) class II molecules and specific VB regions of the T cell receptor. Furthermore, this leads to the activation of both antigen-presenting cells and Т lymphocytes. Ultimately mechanism results in the over-production of inflammatory cytokines and T cells followed by the presentation of autoimmune activated clinical symptoms [6].

The most important superantigens were the Staphylococcus aureus enterotoxins (A, B, C, D and E) and toxic shock syndrome toxin that participate in T cell activation [6]. More than 20 superantigens have been identified in S. aureus strains and at least 80% of the clinical strains of S. aureus harbor a minimum of one superantigen [7].A study in sclerosis(MS) patients examining association of bacterial superantigens with MS exacerbation has been recently carried out [8], therefore, colonization of the nose with enterotoxin A-producing Staphylococcus aureus may play a role in triggering autoreactive CD4+ T cell activation and Multiple Sclerosis exacerbations[9]. The role of S. aureus superantigens in the etiology of MS is currently still unknown. Staphylococcal Enterotoxin Α (sea),Staphylococcal Enterotoxin В (seb),Staphylococcal Enterotoxin C (sec) and Toxic Shock Syndrome Toxin-1 (tsst-1) have been demonstrated to be involved the reactivation of a mouse model for MS ,known experimental autoimmune encephalomyelitis (EAE) [10].

Furthermore, a previous study demonstrated that S. aureus superantigens, used at a nanogram range, were effective in inducing yδT cell activation in a panel of 16 gamma delta T cell clones isolated from MS patients and controls [11].A correlation between superantigens and patients with MS could lead to a better understanding of the etiology of this disease and possibly lead to MS treatment by antibiotic targeting organisms that produce superantigens [9]. The purpose of this study is to determine the prevalence of S. aureus colonization in

non-MS and MS patients, and examine whether there is any correlation between MS patients and *S. aureus* harbouring the superantigen *sea*, *seb*, *and sec* genes.

#### **Materials and Methods**

#### Sample Collection

A total of 200 samples were enrolled in this study. A 100 nasal swab samples were collected from multiple sclerosis patients and 100 from healthy individuals during a period between November 2018 to January 2019 from Baghdad Teaching Hospital / Multiple Sclerosis Clinic in Medical City.

#### **Bacterial Isolation**

All specimens were cultured on 5% sheep blood agar and mannitol salt agar, incubated at 37°C for 24 hours. *Staphylococcus aureus* colonies were identified using phenotypic and biochemical tests like morphology of colony, catalase and coagulase tests [12].

#### **Antimicrobial Sensitivity Testing**

Antibiotic susceptibility test were perfumed for each S. aureus isolate by disc diffusion method according to the clinical laboratory standards institute(CLSI) guidelines [13] against Methicillin (10 µg), Vancomycin (30 µg), Azithromycin(15 µg), Erythromycin (10µg), Tetracycline (10µg), Amikacin (10µg) ,Rifampin (5µg), Penicillin-G(10ug), Meropenem (10ug), Doxycycline (30µg), Sulfamethoxazole/trimethoprim (25 μg), Ciprofloxacin (10 μg), Clindamycin (10 μg), Imipenem (10 μg), Cefotaxime μg), Gentamicin(10μg), Ceftriaxone (10μg), Ampicillin (25 µg), Chloramphenicol (10 ug). All antibiotic disks were provided from Bioanalyse Group Ltd, UK.

#### **DNA Extraction**

The DNA was extracted from pure isolates of *S. aureus* after inoculation in Luria-Bertani broth by Promega genomic purification kit USA according to the company instructions.

#### PCR for Detection of Enterotoxin Genes

Polymerase chain reaction (PCR) was used for amplification and detection of (sea, seb & sec) genes of S.aureus with primers specific to each one were designed and synthesized in Alpha DNA (Canada) (Table 1) and master mix ready to be used obtained from Bioneer, Korea company.

Table 1: Sequences and products of Staphylococcus aureus sea, seb and sec genes

SE gene		$ \begin{array}{ccc} \text{Nucleotide} & \text{sequences} \\ (5' & & 3') \end{array} $	Products bp	References
	F	GGGAACAGCTTTAGGCAATC		
sea	R	ATTTGAATACTGTCCTTGAGC	564	[14]
	F	CCAGATCCTAAACCAGATGAG		
seb	R	TGCAGGCATCATGTCATACC	599	[14]
sec	F	GACATAAAAGCTAGGAATTT	257	[15]
	R	AAATCGGATTAACATTATCC		

PCR amplification of enterotoxin genes was performed in the (Eppendorf, UK) Thermal Cycler under following program: 1 cycle with 94 °C 5 min. for initial denaturation, 30 cycles for denaturation, annealing and extension with 94 °C 2 min, 50 °C 2 min. & 72 °C 1min respectively and finally 72 °C 5 min for final extension [16]. The PCR products were analyzed by (1%) agarose gel electrophoresis [17].

#### Statistical Analysis

The statistical data were analyzed by statistical package (SPSS) ver. (22.0), Chisquare exact test was used to measure significant associations between the frequency of *Staphylococcus aureus* in nasal carriage of MS patients and healthy nasal carriers with enterotoxin genes distribution. P values less than 0.05 were considered statistically significant.

#### Results

# Staphylococcus aureus Colonization and Multiple Sclerosis

Out of 100 nasal swab samples collected from MS patients, 81 (81%) were positive for S.aureus whereas from 100 nasal swab samples collected from healthy controls, there were only 12(12%) positive for S.aureus. Consequently MS patients were most commonly colonized group in this study and there was a statistically significant differences between S. aureus colonization in healthy carriers and MS disease at (p =0.00).On the other hand the relapsing remitting MS patients were most commonly colonized with Saureus 50(61.8%) than newly diagnosed MS group 31(38.2%) with a p-value of (0.092), statistically non significant association between them (Table 2).

Table 2: distribution of Staphylococcus aureus in study groups

Table 2. distribution of Staphytococcus dureus in Stady groups							
Ct1	Samples	S.aureus positive					
Study groups	NO.	NO.	9	<b>6</b>	<i>P</i> -value		
Non-MS group	100	12	1	2	0.00 (HS)*		
Total MS patients	100	81	8	1			
New diagnosed MS group	50	31	38.2				
MS exacerbated group	50	50	61.8	100	0.092 (NS)**		
Total groups	200	93					

<sup>\*</sup> Highly significant; \*\* non significant

## Demographical Distribution and S.aureus Nasal Carriage

According to gender distribution, the ratio of male to female was (1:1.7) includes 64(64%) of patients were female and 36 (36%) were male while in non-MS group there were 54(54%) female and 46(46%) male. In MS patients, a total of *S.aurus* positive was

81(81%) including 52(52%) female and 29(29%) male, while in control non-MS group, out of 12(12%) *S.aureus* positive there were 7(7%) female and 5(5%) male, statistically highly significant differences was observed between *S.aureus* colonization and gender distribution (P value= 0.00) as shown in (Table 3).

Table 3: Distribution S.aureus according to gender

Gender		Multiple sclerosis group		Non-MS co		
		S.aureus		S.aureus		
		(+ve)	(-ve)	(+ve)	(-ve)	
Male	No (%)	29(29%)	7(7%)	5(5%)	41(41%)	0.0 (HS) *
Female	No (%)	52(52%)	12(12%)	7(7%)	47(47%)	0.0 (HS)

Total	No (%)	81(81%)	19(19%)	12(12%)	88(88%)	0.0 (HS)
Total	140 (70)	100(100%)		100(100%)		0.0 (HS)

<sup>\*</sup> Highly significant

#### **Antimicrobial Sensitivity Pattern**

A total of 19 types of antibiotics were tested against *S.aureus* isolates in all study groups according to CLSI. MS patient groups were more resistant to antibiotics (53.3%) than control group (22.8%). Furthermore, within

MS group, the exacerbated MS showed more resistance (60.6%) than newly diagnosed patients (46.3%). In all study groups there is complete resistance to Methicillin (100%) where as the lowest resistance (3%) were reported against Meropenem and Imipenem as shown in (Table 4).

Table 4: Antibiotic resistance pattern of S.aureus isolates.

•	Multi	N. MC		
ANTIBIOTICS	New Dx. MS	R.R. MS.	Total	Non-MS
	No.(%)	No.(%)	No.(%)	No.(%)
Methicillin ME	31(100%)	50(100%)	81(100%)	12(100%)
Vancomycin [VA]	3(9.6%)	5(10%)	8(10%)	0
Azithromycin[AZM]	18(58%)	43(86%)	61(75.3%)	0
Erythromycin [E]	22(71%)	45(90%)	67(82.7%)	8(66.6%)
Tetracycline [TE]	18(58%)	41(82%)	59(72.8%)	4(33.4%)
Amikacin [AK]	17(54.8%)	33(66%)	50(61.8%)	0
Rifampin[RA]	9(29%)	20(40%)	29(35.8%)	0
Penicillin-G [P]	21(67.7%)	47(94%)	68(84%)	12(100%)
Meropenem [MEM]	1(3.3%)	3(6%)	4(5%)	0
Sulfa/Trimthoprim [SXT]	20(64.5%)	45(90%)	65(80.2%)	0
Doxycycline [DOX]	8(25.8%)	26(52%)	34(42%)	0
Ciprofloxacin [CIP]	8(25.8%)	19(38%)	27(33.4%)	0
Clindamycin [DA]	9(29%)	24(48%)	33(40.8%)	0
Imipenem [IPM]	1(3.3%)	2(4%)	3(3.7%)	0
Cefotaxime [CTX]	10(32.3%)	24(48%)	34(42%)	0
Gentamicin [CN]	18(58%)	28(56%)	46(56.8%)	0
Ceftriaxone [CRO]	27(87%)	49(98%)	76(93.8%)	8(66.6%)
Ampicillin [AM]	23(74.2%)	48(96%)	71(87.7%)	8(66.6%)
Chloramphenicol [C]	9(29%)	24(48%)	33(40.8%)	0
Total	46.3% 60.6%			
iotai	53.3	%	53.3%	22.8%

### Conventional PCR Screening for Enterotoxin Genes

produced an amplicon of size 564 bp, 599 bp and 257 bp respectively. Figure (1, 2 and 3).

By PCR essay, the amplification of sequences of seA, seB and seC genes were



Figure 1: Gel electrophoresis of amplified PCR product (564 bp) for sea gene. Lane 1: 100 bp ladder. Lane 2: Negative control; Lanes 3–10: Clinical isolates showing positive result. (1 % agarose, 7 v/cm², 1 hr)



Figure 2: Gel electrophoresis of amplified PCR product (599bp) for seb gene. Lane 1: 100 bp ladder. Lanes 2 – 9: Clinical isolates showing positive result. Lane 10: Negative control, (1% agarose, 7 v/cm², 1 hr)

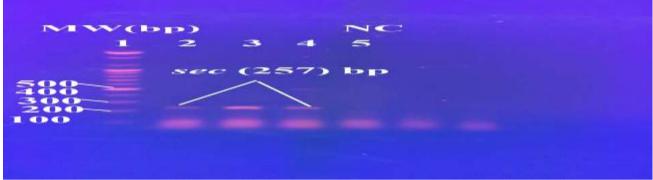


Figure 3: Gel electrophoresis of amplified PCR product (257 bp) for sec gene. Lane 1: 100 bp ladder. Lanes 2 – 4: Clinical isolates showing positive result. Lane 5: Negative control (1 % agarose, 7 v/cm², 1 hr)

The current study reported that, among MS patients the most commonly detected enterotoxin genes were seA 56(69.1%) followed by seB 40(49.4%) while the least prevalent enterotoxin was seC 7(8.6%). Statistically, there was a highly significant differences between the distribution of sea and seb (P=0.00) in MS patient compared with healthy control but non-significant distribution were found in seC (P = 0.138). From overall S.aureus positive in MS groups the distribution of enterotoxins were

36(72%) of se A, 25(50%) of se B & only 4(8%) of se C in relapsing remitting group compared with newly diagnosed MS group that showed 20(64.5%), 15(48.4%) and only 3(9.7%) of se A, se B and se C respectively, whereas no enterotoxin genes were detected in non-MS control group . Significant differences were seen in the distribution of seA, se B and se C gene between MS groups (P=0.00002) compared with controls, (Table 5).

Table 5: Distribution of S. aureus enterotoxin genes

Study groups	S.aureus positive	Gene distribution				P-value
		sea	sea seb sec Total			
	No.(%)	No.(%)	No.(%)	No.(%)	No.(%)	
Control	12(12)	0(0)	0(0)	0(0)	0(0)	
Newly MS	31(38.2)	20(64.5)	15(48.4)	3(9.7)	38(40.8)	0.00002 (HS)*
R.R. MS	50(61.8)	36(72)	25(50)	4(8)	65(43.3)	
Total	93	56(69.1)	40(49.4)	7(8.6)	103(42.4)	
P-value		0.0	0.0	0.138	0.00002	
		(HS)	(HS)	(NS)**	(HS)	

<sup>\*</sup> Highly significant; \*\* non significant

#### **Discussion**

Multiple sclerosis is a multifactorial disease with unknown origin but environmental agents like bacterial and viral infections more participate. *Staphylococcus aureus* superantigens which is considered the most

triggering agent of autoimmunity and included in several autoimmune diseases like multiple sclerosis and rheumatoid arthritis [18]. In multiple sclerosis the presence of *S.aureus* enterotoxins in nasal carriage mostly (sea,seb&sec) considered a potent

factors for induction of autoreactive CD4+ T cells particularly in the myelin protein of CNS hence activation large number of cytokines that finally have a major role in MS severity and development [6,9]. In the present study the frequency of S.aureus nasal colonization in MS patients (81%), particularly in MS exacerbated group was high (61.8%) compared with non-MS healthy carriers which was only(12%) with highly significant differences (P=0.00), this may be due to more contact of these patients with healthcare settings although all MS patients were taken immune activation drugs that enhance immunity which indicates that nasal bacterial colonization with enterotoxigenic S. aureus in MS patients was much more virulent and pathogenic than in non-MS group [8].

The current result in accordance with other study in Iran by *Pakbas et al* who reported that the nasal carriage rate of *S.aureus* in MS patiens & control group was (42%) and (23.3%) respectively [14]. Alternatively a study was conducted in Canada by *Mulvey et al.* revealed a lower prevalence of *S.aureus* in nasal carriage of MS patients (27%) and relatively higher in non-MS group (30%) [9].Regarding gender, this study observed that the frequency of *S.aureus* isolates in MS patients were 29% in males and 52% in females while in control group the percentage of *S.aureus* was 5% males and 7% females.

Concerning MS patients the result of this study coincide with *Sadeghi et al.*,2019,who reported that the frequency of *S.aureus* isolates in MS patients were 23% in males and 76% in females but this result disagreed with the same study [8] that reported the colonization of *S.aureus* in healthy nasal carriers was 56% males and 43% females. Globally *S.aureus* shows high resistance rate to several antibiotics.

The present study revealed that the *S.aureus* isolates in MS patients was (53.3%) multidrug resistance especially in MS exacerbated group (60.6%) than newly diagnosed (46.3%) and control group (22.8%) with highly significant differences between MS patients & non-MS group regarded to resistance pattern. In this study S.aureus isolates in all methicillin groups were resistant to (ME)(100%) indicates that MRSA strains were also distributed in nasal carriage of healthy control [19], as well as there is a high percentage of resistance to ceftriaxone

(93.8%).ampicillin (87.7%). penicillin-G ervthromycin (82.7)(84%),%). sulphamethoxazole/ trimethoprim (80.2 %) azithromycin (75.3%), while in control group in addition to methicillin resistance. S.aureus isolates showed high resistance rate only to penicillin-G(100%), indicating that all strains of S. aureus isolates in MS patients were resistant strains and responsible for multiple drug resistance (MDR) which leads to increase severity of multiple sclerosis(MS), while the S.aureus strain was different in control group in term of limited antibiotic resistance results and so non virulent. This result agreed with Mehrabi et al (2015) [16].

Who reported that resistant S.aureus strains in MS patients was considered as a risk factor for MS exacerbation with high level of resistance, against, tetracycline (80%), ampicillin (72.22%), methicillin (66.66%), erythromycin (66.66%), oxacillin (63.33%) and, sulphamethoxa zole/trimethoprim (61.11%), while more than (90%) of isolates in all study groups were susceptible to vancomycin, meropenem & imipenem which is provided a good regimen for decolonization of S.aureus in nasal carriage of MS patients in order to prevent severity or exacerbation [20].

To the best of our knowledge, it is believed that, it is the first study in Iraq concerning three enterotoxins of *S.aureus* (sea, seb &sec) identified in MS patients. The prevalence of s.aureus enterotoxin genes (sea, seb, sec) was higher in MS patients compared with control group that present no any enterotoxin genes. Statistically significant differences was observed between *S.aureus* harboring enterotoxins in MS patients and Non-MS Carriers (P= 0.00).

Among MS patients the most common prevalent enterotoxins were sea (69.1%) particularly sea in the relapsing remitting group was (72%) and in newly diagnosed MS was (64.5%). In addition the second most common prevalent enterotoxin MS patients was seb (49.4%) with (48.4%) & (50%) in newly diagnosed & R.R. MS patients respectively ,while  $_{
m the}$ least enterotoxins distributed in MS patients was sec (8.6%) with (9.7%) & (8%) in newly diagnosed & R.R. MS patients respectively.

This indicates a statistically significant correlation between *S.aureus* harboring enterotoxin (sea &seb) and MS exacerbation (P= 0.00), however there was no statistically

significant correlation between sec & MS exacerbation (P= 0.138). Similarly other study results by Sadeghi et al. reported that sea was the most common frequent (40%) enterotoxins in MS patients than in nasal carriers (23%) [8], all these results indicate carrying healthy individuals was nonpathogenic strains of S.aureus in nasal passages that considered as a normal commensal flora fight by competing against pathogens, in contrast to S. aureus strains in MS patients were have many virulence factors like enterotoxin genes, more MDR particularly in a stage of relapsing remitting or exacerbated MS that responsible for causing increase sequel. development and pathogenesis of this autoimmune disease.

#### Conclusion

The nasal carriage of *S.aureus* in MS patients particularly in relapsing remitting group was very high compared with healthy individuals that mean the potential risk of *S.aureus* colonization in MS patients was high. High frequency of *S.aureus* harboring enterotoxin genes particularly (*seA*) support the evidence that *seA* play a very significant role in the development and severity of MS. Decolonization regimen of *S.aureus* in nasal carriage of MS. Patients with more effective antibiotics like imipenem to prevent severity or exacerbation of the MS disease.

#### References

- 1. Chabas D, Fontaine B, Lyon-Caen O (2004) Multiple sclerosis review. Orphanet Encyclopedia, 1-6.
- 2. Goldenberg MM (2012) Multiple Sclerosis Review. P T, 37(3):175-84.
- 3. Libbey JE, Cusick MF, Fujinami RS (2014) Role of pathogens in multiple sclerosis. International Reviews Of Immunology, 33(4) 33(4):266-83. DOI: 10.3109/08830185.2013.823422.
- 4. Chowdhary VR, Tilahun AY, Clark CR, Grande JP, Rajagopalan G (2012) Chronic exposure to staphylococcal superantigen elicits a systemic inflammatory disease mimicking lupus. J. Immunol., 189(4):2054-62.
- 5. Proft T, FRASER JD (2003) Review Bacterial superantigens. Clin Exp Immunol 2003; 133:299-306. doi: 10.1046/j.1365-2249.2003.02203.x
- 6. Torres BA, Kominsky S, Perrin GQ, Hobeika AC, Johnson HM (2001) Superantigens: the good, the bad, and the ugly. Exp. Biol. Med., 226(3):164-76.
- 7. Xu SX, McCormick JK (2012) Staphylococcal superantigens in colonization and disease. Front Cell Infect Microbiol, 2 (52): 1-11. DOI: 10.3389/fcimb.2012.00052
- 8. Sadeghi J, Alizadeh N, Oskoueia MA, Laghusi D, Oskoueid DS et al (2019) Frequency of superantigen encoding genes of *Staphylococcus aureus* isolates collected from multiple sclerosis (MS) patients and

- nasal carriers, Microbial Pathogenesis, 127:316-319. doi.org/10.1016/j.micpath.2018.12.010
- 9. Mulvey MR, Doupe M, Prout M, Leong C, Hizon R, Grossberndt A, Klowak M, Gupta A, Melanson M, Gomori A, Esfahani F, Klassen L, Frost EE, Namaka M (2011) Staphylococcus aureus harbouring Enterotoxin A as a possible risk factor for multiple sclerosis exacerbations . Mult. Scler J., 17(4): 397-403. doi: 10.1177/1352458510391343
- Schiffenbauer J, Johnson HM, Butfiloski EJ, Wegrzyn L, Soos JM (1993)
   Staphylococcal enterotoxins can reactivate experimental allergic encephalomyelitis.
   Proc. Natl. Acad Sci. U S A 90(18): 8543-6.
- 11. Shimonkevitz R, Colburn C, Burnham JA, Murray RS, Kotzin BL (1993) Clonal expansions of activated gamma/delta T cells in recent-onset multiple sclerosis. Proc. Natl. Acad Sci., 90(3):923-7.
- 12. Alwan AH, Talak MA (2015) Isolation and characterization of Staphylococcus aureus in spoiled food samples. Int. J. Curr. Microbiol App. Sci., 4(3): 645-651.
- 13. Hindler JA, Schuetz AN (2019) CLSI AST News Update, 4(1):1-21.
- 14. Pakbaz Z, Sahraian MA, Mahmoodi M, Pourmand MR, Sabzi S (2017) Prevalence of sea, seb, sec, sed, and tsst-1 genes of Staphylococcus aureus in nasal carriage and their association with multiple sclerosis. Germs, 7(4):171-7.

- 15. Johnson W M, Tyler S D, Ewan E P, Ashton F E, Pollard D R, Rozee KR (1991) Detection of genes for enterotoxins, exfoliative toxins, and toxic shock syndrome toxin 1 in *staphylococcus aureus* by the polymerase chain reaction. Journal of Clinical Microbiology, 29(3): 426-430.
- 16. Mehrabi F, Asgari A (2015) Resistant Strains of Enterotoxigenic Staphylococcus aureus; Unknown Risk for Multiple Sclerosis Exacerbation. Iran Red Crescent Med J., 17(9):1-7. DOI: 10.5812/ircmj.12596
- 17. Oelkers P (2016) Molecular Biology Laboratory Manual.
- 18. Li J, Yang J, Lu YW, Wu S, Wang MR, Zhu JM (2015) Possible Role of Staphylococcal Enterotoxin B in the

- Pathogenesis of Autoimmune Diseases. Viral Immunol., 28(7):354-9.
- 19. Dağı HT, Fındık D, Demirel G, Arslan U (2015) Detection of Methicillin Resistance and Various Virulence Factors in Staphylococcus aureus Strains Isolated from Nasal Carriers. Balkan Med. J., 32(2):171-5. doi: 10.5152/balkanmedj.2015.150186.
- 20. Buehlmann M, Frei R, Fenner L, Dangel M, Fluckiger U, Widmer AF (2008) Highly Effective Regimen for Decolonization of Methicillin-Resistant Staphylococcus aureus Carriers. Infect Control Hosp Epidemiol. 2008; 29(6):510-6. doi: 10.1086/588201.