



## Determain Kind and Concentration of *Allium sativum* L.Plant Ingredients and its Effect on Isolated Bacteria Causing Urinary Tract Infections

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### Abstract

A total of (190) samples urinary were collected patients with Urinary Tract Infections (UTIs) from the period 2017/11/18 to 2018/2/17, for determine kind and concentration of *Allium sativum* L. ingredients and its effect on growth of the Gram-negative bacteria isolated from urinary tract infections in Al-Alam city/Tikrit Governorate/Iraq. The study was carried out in the laboratories of Al-Alam Hospital and the laboratories of the College of Education for Women, Department of Life Science, chemical analysis by HPLC apparatus showed the plant contains several active ingredients: S-allyl cysteine, γ-glutamylcysteine, Vinyl-4H-1,2-dithiolin (ajoene), di-allyl disulfide and Diallyl trisulfide with concentrations, 5.72%, 9.75%, 53.09%, 9.72%, 12.85% and 10.60% respectively, also appeared many bacteria genera in the samples taken from patients: *E. coli*, *Klebsiella*, *Pseudomonas*, *Proteus*, *Serratia* and *Chromobacterium* with isolates concentration reached for each genus reached, 44.2%, 23.5%, 17.6%, 5.5%, 5.8% and 2.9% respectively, and the current study for aqueous and alcoholic for extracts appeared a clear contrast on inhibition of studied bacteria kinds, as it was alcoholic extract of *Allium sativum* more inhibitory than aqueous extract on the concentration 100% through observation inhibition diameter on the concentration 25, 50 and 100%.

**Keywords:** *Allium sativum*+plant extract+bacteria.

### Introduction

Urinary Tract Infections (UTIs) are the most common diseases, and come after respiratory system diseases, which leads to the death of many people infected with them [1]. (UTIs) infect any part or place of urinary tract and it is caused by many kinds of gram-negative and positive stains which are the causes of the diseases [2]. And the number of people whom infected by it reaches millions, including females and males on different ages, and women are more likely to get the diseases than men [3]. The increased importance of study of bacteria negative gram stain is not only because of its ability on configuration diseases for people who sleep in hospitals but also because of increasing the ability to resist antibiotics, which became a healthy problem scattered in the world [4].

Very few negative bacteria are more common to produce the infection for example kinds do not fermenting glucose such as *E. coli*, *Pseudomonas aeruginosa*, which are considered most prevalent and most infection

after *E. coli*. Because of because it has a ferocity factor characterized by it and giving effective resistance for antibiotic [5]. It has been found 50-60% from these diseases infections in Iran, also it had seen increasing resistance this kind for antibiotics during a few past [6], the negative gram stain bacteria are more kinds finding on urinary tract infection diseases, may be rod shaped, some of them moved and grow on temperature 37°C and some of 30-35°C and its aerobic bacteria negative for oxidase enzyme produce and positive for catalase test except *Shigella dysenteriae* type which exist in water, soil, plant surface and animal body [7].

They found that the largest living organisms that cause UTIs disease are bacteria, including the negative, Enterobacteraceae and Sphingomonadaceae and the positive bacteria, including the Staphylococcus and Spongy [8, 9], this disease considered from medicinal problems which exists on more

countries of the world, either in Iraq it ranks first in between of others bacterial diseases up to 23% [10]. Discovery of antibiotics had important effect on putting ends and outbreaks of diseases and this lead to formation unlimited mounts from it which had role on eradicate the on several type of bacteria [11], but its effect on bacteria began to decline because it had become resistance on this antibiotics and thus become with weak effect on the bacteria, and it has defensive means, and this means be on top of it when this antibiotic used randomly and irregular and in large because quantities bacteria contain plasmid has ability to confrontation the antibiotic and also it has ability to on access to wide distances among the bacteria [12, 13].

The more and bad use of antibiotic be related with fast ability to formation bacterial Strain capable of resistance antibiotic and the relationship will be trivial between bacteria and antibiotic [14], and because of this problem and its side effect the scientists turned to use natural plants to make it treatment for more diseases [15, 16], which sweeps the body Including UTIs disease because medicinal plant has non or limited side effect [17], and with excellent ability on make a physiology change more than industrial and chemical material [18, 19] Treatment with plants and medicinal herbs is one of the method, used throughout the ages in treating various diseases. Many plants have been used for treatment by civilizations, including Indian and Chinese.

This plants has active compounds help human a lot in disposal the diseases [15, 20], compounds as alkaloides, essential oils, glycosides and commarins [21, 22]. Allium sativum plant used around the world because its spicy flavor as one type of spices and this clovers are more uses [23], and it has volatiles oil and sulfide compounds such as allicin, minerals, protein and amino acid [24], in addition of its containing from elements Ca, P, Fe, C and Vitamins which give it an important to Elimination microorganisms and give the body immunity to counter a lot diseases [25] studies confirmed that

*A. sativum* composed of many components of which glucoside sulfure, essential oil, homogeneous mixture of allyle oxide, sulfure, iode and silice and material similar to antibiotics as allicine and garlicine which

they has effective effect against *Streptococcus* bacteria [26], the cellulosic fiber is the main components of *A. sativum* clovers which uses medicinaly with distinctive aroma which is attribute its effect as a wide field antibiotic of microorganisms [27]. In Iraq many researchers they did study about effect of the plant and effect its extracts on the bacteria, [28], showed effective role of aqueous and alcoholic plant extracts on growth of bacteria isolated from tonsils, and there in an important for plant extracts on resistance of lung system bacteria [29], Extracts of *Apium graveolens* and *Trigonella foenum* Working on prevent growth UTIs bacteria and the incidence of the disease [30]. The *A. sativum* plant has been studied and its effect on the growth of negative bacteria has been used in its seeds. The aim of the research to determine kind and concentration of *A. sativum* ingredients and isolation also diagnosis of negative bacteria with Test the effect of plant extracts on the growth of bacteria.

## Material and Methods

About 190 samples of negative gram stain bacteria from patients of UTIs were collected on 18\11\2017 to 17\2\2018, then diagnosed bacteria after their development at the medium blood agar that contain the 5% of the blood of the human and medium MacConkey and EMB sterile autoclave degree 121 degree Celsius and for 15 minutes. As a person depending on the qualities of formal and tests biochemical [31]. Also use the system API 20E make sure of the diagnosis [32].

## Preparing Plants Extracts

*A. sativum* cloves were washed by tap water then with sterilized water and air dried at room temperature, 100gm of cloves milled and used for extraction in 100ml of both hot water and ether alcohol is well known size more for a period of 24 hours in the sifter vibratory and are nominated placed streaming in a centrifugal quickly 3000/5000 cycle/min, and then nominate of new and take streaming and placed in evaporator rotor for getting dry powder then different concentration 25, 50 and 100% prepared from the dried extraction [33].

## Active Ingredients Appreciation

About 10 gm from *A. sativum* put in 50 ml boiled water (90-100°C) for 3 hours then

extracted whattman papers no.1 the extraction collected and put in closed glass tube in order measuring the concentration of active ingredients by High Performance Liquid Chromotography apparatus(HPLC) which supplied by Shimadzu company (Japan) type,LC-10A 2000 supplied with spectrum scale (Spectro photo meter – spd – 10A – UV),Asample size 20µl injected on Fast liquid chramotographic column (LC) with diamention(50×4.6mm I.D) by the injector type (Rheodyn-712) at condition show in

(table 1)and the data recorded by calculator which drewed the pick area and retention time.Astandard solution of *Allium sativum* plant used and sperated by HPLC apparatus and identification the pick area and retention time of standard solution (Table 2 Figure 1 )and comparing it with the pick area and retention time of studed plant sample at the same condition [34]. Concentration of compounds in the sample calculated by the aquation:

Pick area of compounds

$$\text{Conc. Compound in the plant} = \frac{\text{Pick area of compounds}}{\text{Pick area of standard pattern}} \times \text{standard pattern conc.} \times \text{delution factor}$$

Pick area of standard pattern

**Table 1: chromatographics separate condition**

Colum	Mobile Phase	Following rate	Type of detector	Temperature	Fast of recorder paper	Sample Size
Reverse	20mm	1.0 ml/min	Ultra	38C <sup>o</sup>	8 mm/min	3µl
Phase	sodium hydrogen phosphate: 10 mm		violate ray at210 nm			
Column (50×2.0 Mm I.D)	octan sulphonate					50 (v/v)

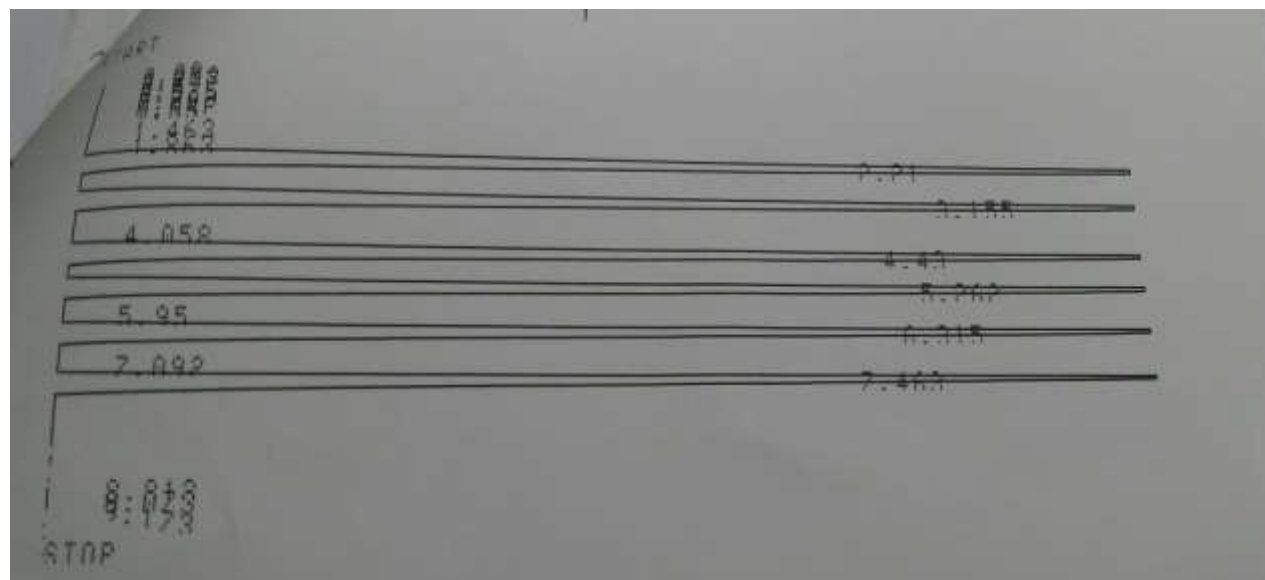
**Bacteria Isolation and Identification**

Bacteria after their development at the medium blood agar that contain the 5% of the blood of the human and medium macConkey and EMB sterile autoclave degree 121

degree Celsius and for 15 minutes. As a person depending on the qualities of formal and tests biochemical [31].Also use the system API 20E make sure of the diagnosis [32].

**Table 2: Compounds, Retention time, Pick area and the concentration of *H. suaveolens***

Compounds	Retention time	Pick area	Concentration (µg/ml)
S-ally cysteine	2.21	157472	25
Y-glutomy cysteine	3.15	270205	25
Allicin	4.43	178054	25
Vinyl-{4H} 1,2dithlin (agoene)	5.26	287693	25
Di-allyl disulfide	6.31	292239	25
Diallyl trisulfide	7.46	336037	25



**Fig.1: Chromatogram HPLC analysis of standard solution of *A. sativum***

## Results and Discussion

Verily Analysis by HPLC apparatus showed existence many compounds in the *A.sativum* cloves as S-ally cysteine, Y- glutomy cysteine, Allicin, Vinyl-{4H} 1,2dithlin(agoene), di-allyldisulfide, Diallyltrisulfide with concentrations were 5.72%, 9.57%, 53.09%, 9.72%, 12.85% and 10,60% respectively (table 3 and Figure 2). Material and active compounds analysis by HPLC apparatus proved its activity in fast on diagnosis this compounds through its ability on calculating the curve with its hight and determine active ingredients in one operation [35].

It is also has the advantage in compare with other methods such as GC by ability on the dealing with non volatail materials including inorganic ions and thermally stable materials [36], results of study agree with [24, 37] whom refers To contain *A. sativum* to Allicin compound and agree with[38] whom confirmed contain this plant to sulfide compounds. After diagnostic tests,number and concentration of isolated bacteria reached 15 ,8,6,2 and 1 isolation by percentage reached 44.2%,23.5%,17.6%,5.8%

and 2.9% for each of *E.coli*, *Klebsiella* , *Pseudomonas*, *Proteus*, *Serratia* and *Chromobacterium* respectively (table 4),the high percentage of *E.coli* due to its own a lot infection factors such as adhesion on Epithelial cells which lining urinary tracts and its own ciliias which assist them adhesion and resistance antibiotics by proudcing the Hymolycin and its ability to growing fastly as it has short generation time [39] *Klebsiella* has factors assist them to occurrence the disease as portfolio protect them from unsuitable conditions and resist phagocytosis process besides of ciliias ,and which givt them another chance to cause disease [40].

While appearance *Serratia* in this percentage agree with [41] *pseudomonas* were and *pseudomonas* is coliform shaped it exists in the form single or pairs or as chains [42] while *proteus* spherical shape and moving bacteria [43] its percentage disagree with [44], the last bacteria species *Chromobacterium* appeared by 2.9% percentage is the most kind cause the disease and which are considered natural flora existing in human and animal intestines [3].

Table 3: Ingrdients, Retention time, pick area, Concentration of *A.sativum*

S	Ingredients	Retention time	Pick area	Concentration µg/ml	Percentage (%)	Dilution
1	S-allyl cysteine	2.17	46894	314	%5.72	50
2	Y-glutomyl cysteine	3.12	64462	438,32	%9.57	50
3	Allicin	4.39	429028	2909,7	%53.09	50
4	Vinyl-{4H}1.2dithlin (agoene)	5.18	80085	533,2	%9.72	50
5	di-allyldisulfide	6.26	102374	704,26	%12.85	50
6	Diallyltrisulfide	7.38	80935	581,09	%10.60	50

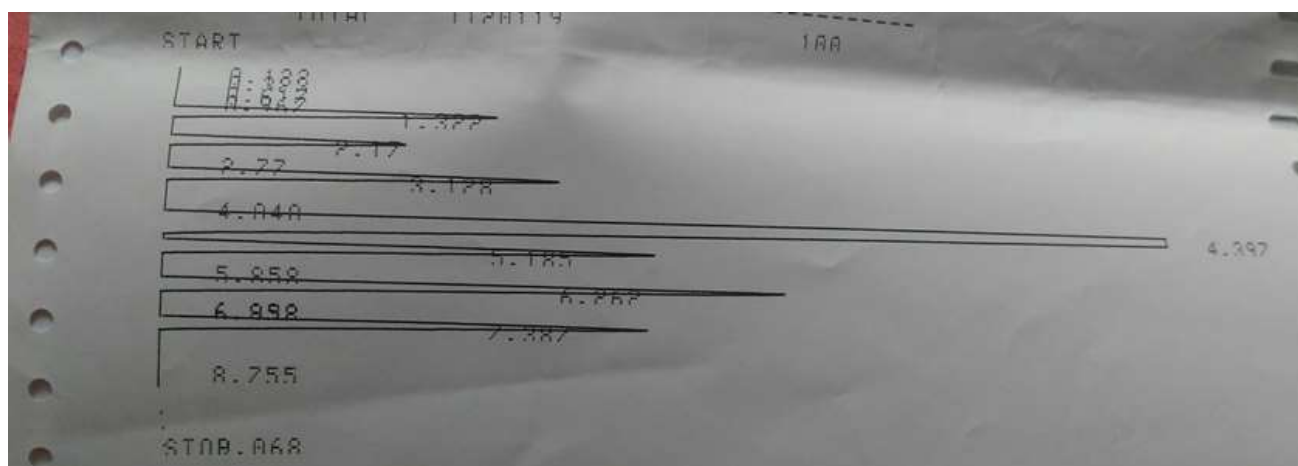


Fig. 2: Retention time and Pick area of *A.sativum* plant ingredients

Table 4.kinds of isolated bacteria

S	Bacteria genus	No.Isolation	Percentage (%)
1	<i>E.coli</i>	15	44.2
2	<i>Klebsiella</i>	8	23.5
3	<i>Pseudomonas</i>	6	17.6
4	<i>Proteus</i>	2	5.8
5	<i>Serratia</i>	2	5.8
6	<i>Chromobacterium</i>	1	2.9

The present study of aqueous and alcoholic extracts of *A.sativum* appeared a clear contrast on bacteria inhibition table 5 and *Pseudomonas* bacteria were most affected than other isolations, and alcoholic extract were more inhibition than aqueous extract by observation inhibition diameter ,highest inhibition diameter by concentration 25% ,50% and 100% reached (13,16 and 20 mm) respectively. The same study showed that *Pseudomonas* more affected by different concentration of aqueous extract than other bacteria 25%, 50% and 100% concentration gaved high inhibition diameter reached (12, 14 and 17 mm) respectively, while *E.coli* bacteria were less affected at the same concentration of both alcoholic and aqueous extraction by inhibition diameter reached (6, 8 and 10 mm) and (5, 6 and 8 mm) respectively. The wide-ranging antagonism of *A. sativum* due to its containing sulfide

compounds such as Allicin, Thiosulfinates [45, 46] whereas allicin works to Partial inhibition of formation the DNA and protein and total inhibition of RNA [47] as that presence of allicin substance has effect on enzymes oxidate [48] In addition to existence sulfide compounds such as Dimethyl disulfide, Methyl methey ethiosulfphonate, diallylsulfide, this compounds has effectiveness against for microorganism growth in addition to non sulfide compounds such as Vitamin B, proteins, Fe, Soaponins and Flavonoids [49] the resultas agree with [50] whom refer to effectiveness of aqueous and alcoholic extracts against isolates negative and positive gram stain bacteria, also agree with [51] whom showed effectiveness aqueous and alcoholic extracts of *A.sativum* against some kind of *Proteus*, *E. coli*, *Pseudomonas* bacteria.

**Table 5.Effect aqueous and alcoholic of *A.sativum* extracts on bacteria isolates**

Extracts Isolates bacteria	Aqueous			Alcoholic		
	Concentrations (%)			Concentrations (%)		
	25	50	100	25	50	100
<i>E.coli</i>	5	6	8	6	8	10
<i>Klebsiella</i>	6	10	12	8	10	14
<i>Proteus</i>	6	8	10	10	12	15
<i>Serratia</i>	6	7	8	6	8	11
<i>Pseudomonas</i>	12	14	17	13	16	20
<i>Chromobacterium</i>	8	12	14	10	14	16

## References

- Leppert PC, Peipert JF (2003) Primary care for women. 2nd ed. Wolters Kluwer/ Lippincott. Williams and Wilkins. Philadelphia, USA, 1: 496-499.
- Tangho EA, McAminch JW (2008) Smith's general urology, 17th ed. McGraw- Hill companies, Inc. USA, 193-196.
- Ahmad W, Jamshed F, Ahmad W (2015) Frequency of Escherichia coli in Patients with community acquired urinary tract infection and their resistance pattem against some commonly used antibacterials. J.Ayub. Med. Coll. Abbottabad., 27 (2): 333-337.
- Sepehri G, Nejad HZ, Sepehri E, Razban S (2010) Bacterial profile and antimicrobial resistance to commonly used antimicrobials in intra-abdominal infections in two teaching hospitals. Am J. Applied Sci., 7: 38-43.
- Karlowsky JA, Draghi DC, Jones ME (2003) Surveillance for antimicrobial susceptibility among clinical isolates of *Pseudomonas aeruginose* and *Acinetobacter baumannii* from hospitalized patients in the United States, 1998 to 2001, Antimicrob Agents Chemother, 47: 1681-8.
- Khalil H, Soltani R, Afhami S, Dashtikhavidaki S, Alijani B (2012) Antimicrobial resistance pattern of gram-negative bacteria of nosocomial origin at a teaching hospital in the Islamic Republic of Iran. East Mediterr Health Journal, 18(2): 172-7.
- Brook GF, Carroll KC, Butel JS, Morse SA (2007) Jawetz, Melnick, and Adelbergs. Medical microbiology. 24th ed. MeGra-Hill. U.S.A.
- Verle VA, Jefferson BM, Arnold Joseph MF, Januario DV, Sonu B (2015) Predict Urinary Tract Infection and to Estimate Causative Bacterial Class in a Philippine Subspecialty Hospital. Singapore. J. Neph. Ther., 5(2):2-6.

9. Emamghorashi F, Shohreh F, Mehdi K, Shadokht R, Maryam H (2011) The Prevalence of O Serogroups of Escherichia coli strains causing acute urinary tract infection in children in Iran. Saudi. J. Kidney Dis., Transpl., 22(3): 597-601.
10. AL-karawyi AM, AL-Jubouri SA, Alasadiy YD (2013) Molecular Detection of AmpC Family Genes Encoding Antibiotic Resistance among Escherichia coli isolated from Patients with Urinary Tract Infection (UTI) in Najaf Hospitals. Vet. Med. Sci., 4(1): 152-161.
11. Ritchie ND, Mitchell TJ, Evans TJ (2012) What is different about serotype 1 pneumococci 2 Future microbial, 7(1): 33-46.
12. Sani I (2014) Application of Medicinal Plants to Overcome Antibiotic Resistance in Some Selected Multi-Drug Resistant Clinical Isolates. Res. and Rev. J. Pharm. and Phyt. 2(4).
13. Dijkshoorn L, Ursing BM (2000) Strain, Clone and species. Comments on three basic concepts of bacteriology. J. Med. Microbiol., 49: 397-401.
14. Keren R, Chan E (2002) Ametaanalysis of randomized cotrolled trials comparing short and long course antibiotic therapy for urinary tract infection in children-pediatrics. 109: 70.
15. Zahin M, Aqil F, Khan MSA, Ahmad I (2010) Ethnomedicinal plants derived antibacterial and their prospects. Res. Signpost 37 /66/ (2) Fort P.O. Trivandrum-695023. Kerala, India.
16. Sibanda T, Okoh AI (2007) The challenges of overcoming antibiotic: plant extract as potential sources of antimicrobial and resistance modifying agents. African J. of Biotech. 6(25): 2886-2896.
17. Essawi T, Srouf M (2000) Screening of palestinian medicinal plants for antibacterial activity. J. Ethnopharmacol., 70: 343-349.
18. Manges AR, Johnson JR, Foxman B (2001) Widespread distribution of urinary tract infections caused by a multidrug-resistant Escherichia coli clonal group N. Engl. J. Med. 342: 355.
19. Andrews SJ, Brooks PT, Hanbury D (2002) Ultrasonography and abdominal radiography versus intravenous urography in investigation of urinary tract infection in men: Prospective incident cohort study. BMJ., 324-454.
20. AL-Naimy EH, Al-Lihibi RK, Majeed SM, Al-Ani RS (2012) Antimicrobial and cytotoxic effects of the kiwi fruit and pomegranates on tumor cell line (L20B, RD). The Iraq J. of Agricultur Sci., 43(1): 147-167.
21. Gawish A, Mohammed N, El-Shennawy G, Mohammed H (2013) An investigation of type 3 secretion toxins encoding-genes of Pseudomonas aeruginosa isolates in a university hospital in Egypt. J. of Microbio. And Infec. Dise., 3 (3): 116-122.
22. Scott JAG, Brooks WA, Peiris JS, Holtzman D, Mulhollan EK (2008) Pneumonia reseach to reduce childhood mortality in the developing world. J. Clin. Invest, 118(4): 1291-1300.
23. Major Food (2013) Agriculture Commodities and Producer- countries by commodity "Fao. Org.
24. Tsao SM, Yin MC (2001) Invitro antimicrobial activity of four diallyl sulphides occurring naturally in garlic and chinese leek oils. J. Med. Microbial, 50:646-649.
25. Cruichshank R, Duguid JP (1999) The practic of medical microbiology. 12th ed. Vol.11, Edinburgh; churchillliving stone, London, New York.
26. Breidenstein EBM, De la Fuente-Nuñez C, Hancock REW (2011) Pseudomonas aeruginosa: all roads lead to resistance. Trends Microbiol., 19: 419-426.
27. McFerren M (2007) " Garlic: what we know and what we don't know". Arch. Intern. Med. 167(4): 325-6.
28. Hussain SHA (2001) Biological effect of some plant extracts and its active components on growth three kind of organism isolated fron tonsilit. Msc Thesis, Coll. Edu. Musol. Univ. Iraq.
29. Al-Saady HAH (2005) Immunological study of desolve enzyme for protein extracted from Proteus mirabilis bacteria isolated from Urologic infections. Msc. Thesis, Coll. Sci. Mustanseryia Univ. Iraq.
30. Al-Wandawy Ash, Al-Samrray M, Al-Janabi SA, Mohhamad H (2002) Combination among Celery seeds, Fenugreek and plant extracts on



- prevention growth and morphology of bacteria causes urinary tract infection, 2th local conference of Sci. Dept. Coll. Sci. Mustanseryia Univ.Iraq.
31. Leboffe MJ, Pierce BE (2011) Aphotographic Atls of the Microbiology Laboratory, 4<sup>th</sup> ed, USA, 58-64, 74-78, 96-99, 153.
  32. Atlas RM (1995) Principle of microbiology. 1st ed. Mosby-Year book, Inc. St. Louis. USA.
  33. Al-DawsaryAWJ (2012) Study effect of aqueous stracts of Garlic and Ginger plants on some physiological and Biochemical properties of broiler and local Iraqi sheeps.Anbaar J. Vet. Sci., 5 (1).
  34. Nishizawi H, Okimura S, Abe Y (1991) Application liquid particle extraction to the purification of glycyrrhizin.Chemical Pharm. Bull., 39: 696-971.
  35. Al-Hedwony AKH (2004) Effect Fertilization and foliar application of some nutrient elements on quantitative and qualitative of some active compounds in seed two kind of (Trigon olla foenum) Ph.D Theses. Col. Agric. Univ. Baghdad, Iraq.
  36. Settle FA (1997) Handbovk of instrumental techniques for analytic chemistry. N.J. Prentic Hall PTR, 945.
  37. Ankri S, Mirelman D (1999) Antimicrobial properties of allicin from garlic, Microbes and Infect, 2: 125-129.
  38. Lawson LD, Wood SG, Hughes BG (1991) HPLC analysis of allicin and other thiosulfinates in garlic clove homogenates. Planta Med., 57(3): 263-70.
  39. Stamm WE, Hooton T M, Johnson JR (1999) Uurinary tract infection: From pathogenesis to treatment. J. Infect. Dis., 159:400.
  40. Jehad M, Abudaia A, Al-Aaly A, Roberto DC (200) UTI in childhood. Saudi Med. J., 21 (8): 711-715.
  41. Al-Abdly YA (2010) Exraction and purification Cathshin compound from green tea (Camellia sinesis) and its maxiture effect on UTIs bacteria. Msc. Thesis, Coll. Sci. Mustanseryia Univ.Iraq.
  42. Delost MD (1997) Introduction to Diagnostic Microbiology, Textbook and workbook. Mosby-Year Book, Inc., St-Louis, Missouri, USA.
  43. Al-Taai H, Rasheed R (2016) Bacteriological comparison Between Pseudomonas aeruginosa and Klebsiella Pneumonia isolated from Different infection Sources, 12 (1): 167.
  44. Jarvis A, Scolnik D (2000) Clinical perspective on diagnosis of urinary tract infection in children. Can. J. Emergency. Med., 2: 201-209.
  45. Hughes BG, Lawson L (1991) Antimicrobial effects of allium sativum and alliumcepa, garlic compound and commercial garlic supplement product. Phytother Res, 5: 154-158.
  46. Lawson LD, Wood SG, Hughes BG (1991) HPLC analysis of allicin and other thiosulfinates in garlic clove homogenates. Planta Med, 57(3): 263-70.
  47. Retty AF, Danil FS, Aice SW (2007) Balley and Scott's of Diagnostic Microbiology, 12th ed. Press, Houston, Texas.
  48. Sovova M, Sova PM (2004) Pharmaceutical importance of Allium sativum L. 5. Hypolipemic effects in vitro and in vivo (in Czech). Ceska. Slov. Farm., 53(3): 117-23.
  49. Bisset NG, Wichtl M (2001) Herbal Drugs and Phytopharmaceuticals. 2<sup>nd</sup> ed., Medpharm Scientific Publishers, Germany, ISBN: 0-8493-1011-3 350-420.
  50. Ameen AA, Hind AS, Ali TA (2015) Invitro”Antibacterial characteristics of garlic plant extract on some pathogenic bacteria kind isolated from burns unit.Thiqaar J. Sci., 5(3):17-21.
  51. Ishrat R, Akhund S, Abro H (2008) Antimicrobial potential of seed extract of Raphanus sativum, Pak. J. Bot., 40(4): 1793-1798.