



Molecular Detection of *Lysinibacillus Fusiformis* Isolated from Milk Samples of Cow in Iraq

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Abstract

Raw milk infected by bacteria can start from exceptional resources: air, milking devices, feed, soil, dungs and hays. The strategies of nourishing and lodging cows may impact the germs pleasant of milk. Bacterial contaminants can cause infection, or deterioration of drain and its auxiliary items and the destructive impacts on the grade and security of dairy items as a result of oxygen consuming spore-forming bacteria got from crude drain were characterized by isolation of *Lysinibacillus fusiformis* from 90 milk samples of cow at 21.61%, the bacteria were identified by routine bacteriological methods and Molecular distinguishing proof of the confines was carried out by 16S rRNA sequencing and the bacterial confines were taxonomically classified as *Lysinibacillus fusiformis*. The groupings were stored in NCBI GenBank with the accession number KY038703, KF916675. and KF916675.1 with identity of 100%, 96 and 99% respectively.

Keywords: Raw milk, *Lysinibacillus fusiformis*, 16S rRNA PCR sequencing.

Introduction

Milk can be considered as a huge supply of microorganisms because milk contains high level of supplements makes it a particularly reasonable development medium for different microscopic organisms. In truth, these microorganisms can accomplish more populace densities taking after contamination amid drain preparing in dairy ranches and in the dairy industry [1].

In truth, it is exceptionally critical to preserve them in reasonable conditions (heat degree and wetness), that maintain a strategic distance from the expansion of the microflora display on their surfaces [2]. Indeed, in commercially sterilized drain, the decay caused by *Bacillus* species has been detailed, in spite of the fact that usually generally caused by thermostable proteolytic and lipolytic proteins or by re-contamination of the sterilized drain amid filling [3].

In arranging to test the microbial groups colonizing books and documented facts, various strategies are handy. In such methodologies can be isolated essentially to two assorted procedures: the culture-dependent strategies (based on the development of microorganisms on particular

microbiological media) and the culture independent methods (based on the extraction of nucleic acids and their examination). *Lysinibacillus fusiformis* is Gram-positive, rod-shaped microbes non motile having a place to the family Bacillaceae [4,5]. The bacteria have a type of peptidoglycan in their cell wall and many researchers' studies its pathogenicity [6].

Lysinibacillus fusiformis separated from other near of Bacillacea since of the presence of lysine and aspartate in the peptidoglycan of the cell wall [7]. The bacteria resist many metals like inmercury-contaminated sites and boron [8,9].

Lysinibacillus fusiformis is pathogenic to human [10], other researcher studies its biosurfactant activity against biofilm formation of *Escherichia coli* and *Streptococcus mutans* [11], else it increased the biofilm of *Bacillus subtilis* by the overabundance of hypoxanthine inside *B. subtilis* cells, which may prompt cell stress and death [12]. This study aimed was isolating of *Lysinibacillus fusiformis* from raw milk of cow and detect it by 16 serene.

Material and Methods

Sampling

Crude drain tests (90) were aseptically collected from dairy ranches at geologically distinctive areas within the locale of Baghdad from a late summer/autumn and a winter.

Bacterial Strain Isolation and Identification

Milk samples were collected in sterile tubes, 0.1ml of every raw milk sample turned into streaked out on blood agar plates (Oxoid) and incubated at 37C with 5% CO₂ for 24 h. Spore-forming bacteria were isolated from media. Each colony was purified by sub-culturing. Isolates were preserved in 20% glycerol at -80 °C.

Biochemical and Molecular Characterization of Isolate

Based on the isolated colony and its morphological and biochemical characteristics were explored. The biochemical characterization of this strain, referred to as *Lysinibacillus fusiformis*.

DNA Extracted

DNA extract was done as described by [13].

PCR and DNA Sequencing

The 16S rRNA may be a well-conserved, all inclusive bacterial genewidely utilized to distinguish contrasts among bacteria. The PCR amplification was performed using universal primers f F₁₆M (5'-AGAGTTTGGATCCTGGCTCAG-3') and R₁₆M (5'-GGTTACCTTGTTACGACTT) – the procedure done as (Coorevits et al 2008). All isolates were sent to (NCBI) and BioEdit program.

Results

Bacterial Isolation

Three isolated bacteria classified as *Lysinibacillus fusiformis* out of 90 raw milk samples in 21.61%.

Characterization of Bacterial Strains

We screened a collection of 90 raw milk samples obtained from different sampling sites in Baghdad in order to distinguish microbes that are found in raw milk. The isolated bacteria from raw milk were distinguished based on biochemical test as *Lysinibacillus fusiformis* as shown in Table 1

Table 1: Some Biochemical test on *Lysinibacillus fusiformis*

Biochemical test and gram stain	<i>Lysinibacillus fusiformis</i>
Gram stain	Negative
Motility	+
Oxidase	+
catalase	+
Citrate utilization	v
Indole	-
Methyl red	-
Urea	+
Hydrolysis of esculin	-
Voges Proskauer's	-
Phenyl alanine deamination	+
Glucose	-
Lactose	-

+indicates positive result, - indicates a negative result, V indicates variable

Evaluation of Bacterial Detection and Identification by 16SrRNA and Sequencing

Lysinibacillus fusiformis nucleotide sequence was submitted to GenBank in NCBI with the

accession number the PCR-sequenced done with the Maxime PCR PreMix kit (i-Taq) 20µl rxn with 100%, 96% and 99% identified as shown in Table 2. The 16s rRNA was amplified (Fig. 1) and subjected to purification and sequencing.

Table 2: Identified bacteria based on sequencing and their similarity with GenBank

Bacteria	Sequence ID	Gen Bank accession no.	% similarity/GenBank closest relative
<i>Lysinibacillus fusiformis</i>	KY038703.1	JN1425	100%
	KF916675.1	JN1435	96%
	KM280032.1	JN915	99%

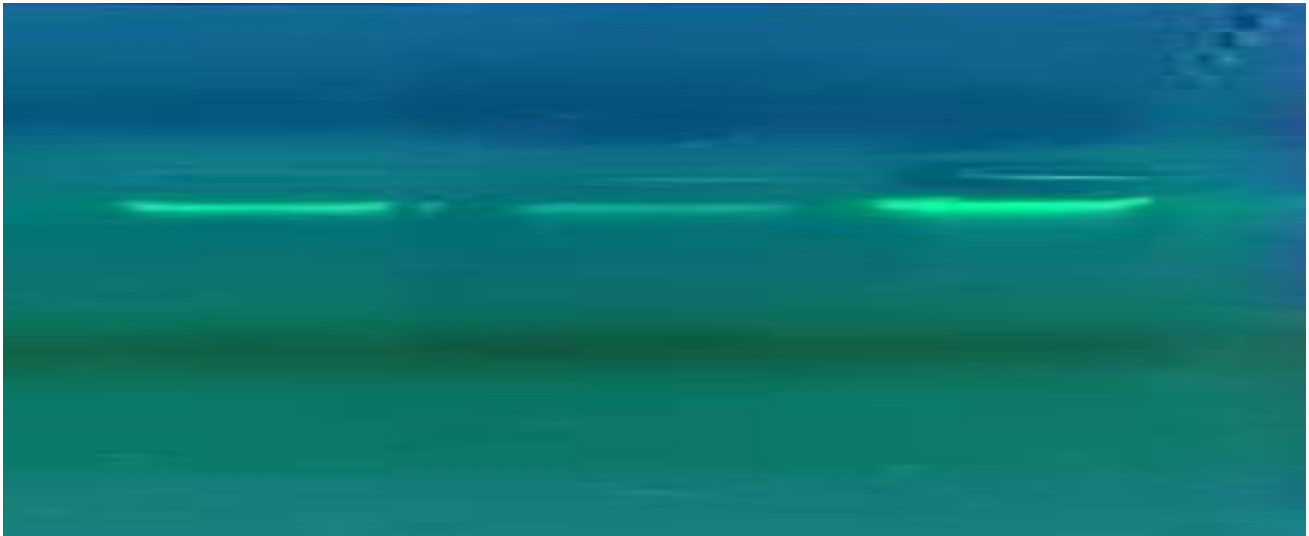


Figure | 1: Gel electrophoresis of genomic DNA extraction from bacteria, 1% agarose gel at 5 vol /cm for 1:15 hour

The homology to *Lysinibacillus fusiformis* DNA sequencing demonstrate 100%,96% and 99% of isolates as shown (Fig.3,4 and 5

respectively). The sequence (approximately 1620, 1817and 1315 bp, respectively) (Fig. 2).

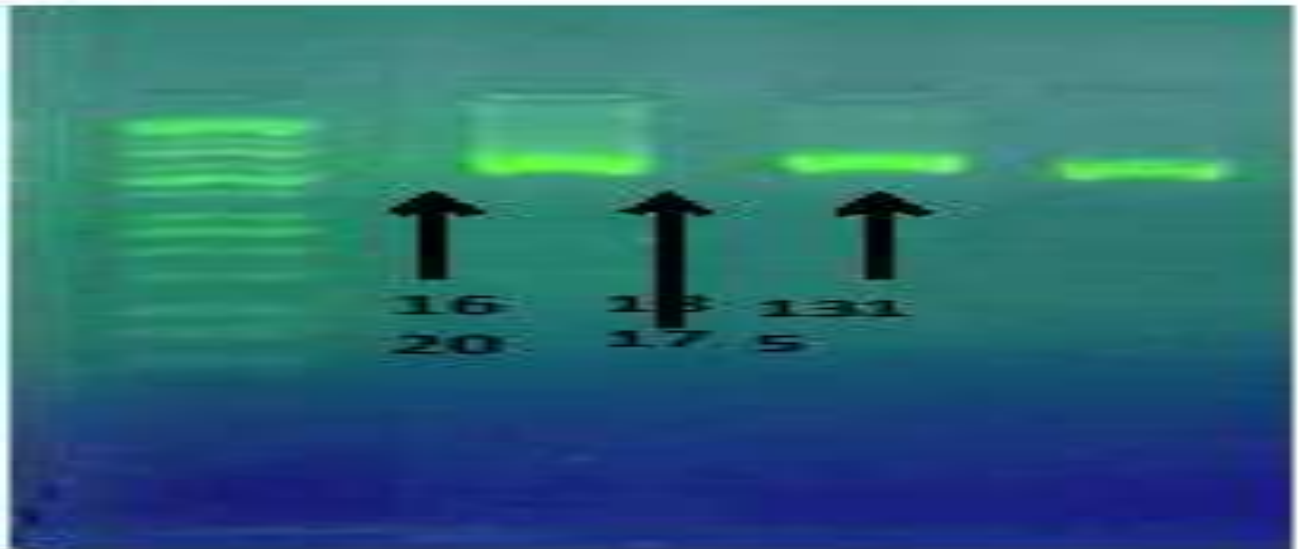


Figure 2: PCR product the band size 1600 bp. The product was electrophoresis on 2% agarose at 5 volt/cm². 1x TBE buffer for 1:30 hours. N: DNA ladder (100)

Score	Expect	Identitas	Gaps	Strand
1620 bits(1796)	0.0	898/898(100%)	0/898(0%)	Plus/Plus
Query: 1	ACATGCACTCGAGCGAACAGAGAAAGGAGCTTGCTCCTTCGACGTTAGCGCGGACGGGTC			60
Subject: 1	ACATGCACTCGAGCGAACAGAGAAAGGAGCTTGCTCCTTCGACGTTAGCGCGGACGGGTC			60
Query: 61	AGTAAACAGTGGGCAACCTACCTTATAGTTTGGGATAACTCCGGGAACCGGGCTAATA			120
Subject: 61	AGTAAACAGTGGGCAACCTACCTTATAGTTTGGGATAACTCCGGGAACCGGGCTAATA			120
Query: 121	CCGAATAATCTSTTTCACCTCATGGTGAACACTGAAGACGGTTCGGCTGTCCGTATA			180
Subject: 121	CCGAATAATCTSTTTCACCTCATGGTGAACACTGAAGACGGTTCGGCTGTCCGTATA			180
Query: 181	GGATGGGCCCGCGCGCATTAGCTAGTTTGTGAGGTAAACGCTCACCAAGCGGACGATGC			240

Figure 3: The sequence of *Lysinibacillus fusiformis* that explain similarity 100% of the first isolate as shown in table 2

Score	Expect	Identities	Gaps	Strand
1817 hits(2014)	0.0	1067/1107(96%)	0/1107(0%)	Plus/Plus
Query 1	ATGCAAMTCCGACCGACA GAAGCGAGCTTCCTCCCGGATGTTAAGGACCGACCGATTGTT	60		
Subject 18	ATGCAAGTCCGACCGACA GATGGAGCTTCTCTCCTGA TGTETACCGCCGACCGATTGTT	74		
Query 61	AACAAGTGGGTAACCTGCTGTAAAGCTGGGATAGCTCCGGAACCGGACCTAAATCGG	120		
Subject 76	AACAAGTGGGTAACCTGCTGTAAAGCTGGGATAGCTCCGGAACCGGACCGCTAAATCGG	134		
Query 181	GATAGTTCCTTAAACCGCATGTTCCAAAGATGAAAGACGGTTCCGCTGTACTTACAG	120		
Subject 193	GATAGTTCCTTAAACCGCATGTTCCAAAGATGAAAGACGGTTCCGCTGTACTTACAG	174		
Query 191	TGGAACCCGCGGCTATTACCTAGTTGCTGGTAAAGCTCCACCGGACCGACCGGAC	240		
Subject 193	TGGAACCCGCGGCTATTACCTAGTTGCTGGTAAAGCTCCACCGGACCGACCGGAC	254		
Query 241	CGGCACTTGAAGCTTATCGGACCGACCTGGGCTTAAAGACCGGCTTAACTGCTACCG	300		
Subject 255	CGGCACTTGAAGCTTATCGGACCGACCTGGGCTTAAAGACCGGCTTAACTGCTACCG	314		

Figure 4: The sequence of *Lysinibacillus fusiformis* that explain similarity 96% of the second isolate as shown in table 2

Score	Expect	Identities	Gaps	Strand
1315 hits(1458)	0.0	738/744(99%)	0/744(0%)	Plus/Plus
Query 1	AGCGGCGGACCGGTTGAGTACACGTTGGTAACTGCCTGTAAAGATGGGATAACTCCGGG	60		
Subject 41	AGCGGCGGACCGGTTGAGTAAACGTTGGTAACTGCCTGTAAAGATGGGATAACTCCGGG	100		
Query 61	AAACCGGAGCTAATACCGGATAGTTCCCTTGAACCGCATGGTTCAAGGATGAAAGACGGTT	120		
Subject 101	AAACCGGAGCTAATACCGGATAGTTCCCTTGAACCGCATGGTTCAAGGATGAAAGACGGTT	160		
Query 121	TCGCTGTCACTTACAGATGACCCGCGGCGCATTAGCTAGTTGGTGGGTTAATGGCTCA	180		
Subject 141	TCGCTGTCACTTACAGATGACCCGCGGCGCATTAGCTAGTTGGTGGGTTAATGGCTCA	220		

Figure 5: The sequence of *Lysinibacillus fusiformis* that explain similarity 99% of the third isolate as shown in table 2

Discussion

The genus *Lysinibacillus* contain more species and the bacteria *L. fusiformis* is one of the most recognized species in this genera [14]. (Priest et al 1988).The bacterial strain *Lysinibacillus fusiformis* was first isolated in 1901 from crop, *Beta vulgaris* cited by [15].

The bacteria confined from crude drain test among several bacterial colonies and appeared capacity to spoilage the raw milk as well as isolated from different environmental sources like exfoliating cream , tamarind seed (*Tamarindus indica*,cow dung,raw milk and dairy farms, waste material and industrial effluent etc. [15,16,17,18,3,19,8]. The bacteria resist many metal and antibiotic and it cause a disease to human such as tropical ulcers and dermal respiratory infections [8,10].

The aim of this research was to evaluated the spore forming bacteria in untreated milk and the influence of season on bacterial isolate, the prevalence of *Lysinibacillus fusiformis* isolation was 21.61% this percent indicate that the raw milk contaminated with bacteria Besides, as changes in cultivate operational management will influence the

high-impact spore-forming microbiota in drain, conceivably indeed select for as yet unknown species, uncommon sharpness on microbial milk quality is required [3]. Moreover Pasteurization and refrigeration of milk very necessary to decreased contamination which lead to milking spoilage that's agree with [20], all isolated obtained during spring and early summer that indicate the bacteria prefer moderate temperture for its multiplication .

The bacteria was isolated and identified by culturing characteristic, biochemical test and confirm diagnosis via 16S rRNA gene sequence Besides, 16S rRNA gene sequencing has been connected to affirm the bacterial species distinguishing proof data[21,22] .

The 16sRNA gene sequencing of *Lysinibacillus fusiformis* was compared with sequences in the NCBI GenBank database with pairwise similarity values of 100%, 96% and 99 %, respectively in the present study one *Lysinibacillus fusiformis* obtaine 96% similarity which needs to be further characterized.

To the authors' knowledge, this is the first documentation of *Lysinibacillus fusiformis*

bacteria, isolated from raw milk in Baghdad, Iraq.

References

- Bartoszewicz M, Hansen BM, Swiecicka I (2008) The members of the *Bacillus cereus* group are commonly present contaminants of fresh and heat-treated milk. *Food Microbiology*, 25(4): 588-596.
- Kraková L, Šoltys K, Otlewska A, Pietrzak K, Purkrťová S, Savická D, Demnerová K (2018) Comparison of methods for identification of microbial communities in book collections: Culture-dependent (sequencing and MALDI-TOF MS) and culture-independent (Illumina MiSeq). *International Biodeterioration & Biodegradation*, 131: 51-59.
- Coorevits A, De Jonghe V, Vandroemme J, Reekmans R, Heyrman J, Messens W, Heyndrickx M (2008) Comparative analysis of the diversity of aerobic spore-forming bacteria in raw milk from organic and conventional dairy farms. *Systematic and Applied Microbiology*, 31(2): 126-140.
- Mulet M, David Z, Nogales B, Bosch R, Lalucat J, Garcia-Valdes E (2011) *Appl. Environ. Microbiol.*, 77: 1076-1085. doi:10.1128/AEM.01741-10
- Nam YD, Seo MJ, Lim SI, Lee SY (2012) *J. Bacteriol.*, 194: 59-88. doi:10.1128/JB.01485-12
- Divakar K, Prabha S, Gautam P (2017) Purification, immobilization and kinetic characterization of GxSxG esterase with short chain fatty acid specificity from *Lysinibacillus fusiformis* AU01. *Biocatalysis and Agricultural Biotechnology*, 12: 131-141.
- Ahmed I, Yokota A, Yamazoe A, Fujiwara T (2007) Proposal of *Lysinibacillus boronitolerans* gen. nov. sp. nov., and transfer of *Bacillus fusiformis* to *Lysinibacillus fusiformis* comb. nov. and *Bacillus sphaericus* to *Lysinibacillus sphaericus* comb. nov. *International Journal of Systematic and Evolutionary Microbiology*, 57(5): 1117-1125.
- Gupta S, Goyal R, Nirwan J, Cameotra SS, Tejoprakash N (2012) Biosequestration, transformation, and volatilization of mercury by *Lysinibacillus fusiformis* isolated from industrial effluent. *J. Microbiol. Biotechnol.*, 22(5): 684-689.
- Raja CE, Omine K (2012) Characterization of boron resistant and accumulating bacteria *Lysinibacillus fusiformis* M1, *Bacillus cereus* M2, *Bacillus cereus* M3, *Bacillus pumilus* M4 isolated from former mining site, Hokkaido, Japan. *Journal of Environmental Science and Health, Part A*, 47(10): 1341-1349.
- Wenzler E, Kamboj K, Balada-Llasat JM (2015) Severe sepsis secondary to persistent *Lysinibacillus sphaericus*, *Lysinibacillus fusiformis* and *Paenibacillus amylolyticus* bacteremia. *International Journal of Infectious Diseases*, 35, 93-95.
- Pradhan A K, Pradhan N, Sukla LB, Panda PK, Mishra BK (2014) Inhibition of pathogenic bacterial biofilm by biosurfactant produced by *Lysinibacillus fusiformis* S9. *Bioprocess and biosystems engineering*, 37(2): 139-149.
- Gallegos-Monterrosa R, Kankel S, Götze S, Barnett R, Stallforth P, Kovács ÁT (2017) *Lysinibacillus fusiformis* M5 induces increased complexity in *Bacillus subtilis* 168 colony biofilms via hypoxanthine. *Journal of bacteriology*, 199(22): e00204-17.
- Heyndrickx M, Vauterin L, Vandamme P, Kersters K, De Vos P (1996) Applicability of combined amplified ribosomal DNA restriction analysis (ARDRA) patterns in bacterial phylogeny and taxonomy. *Journal of Microbiological Methods*, 26(3): 247-259.
- Priest FG, Goodfellow M, Todd C (1988) A numerical classification of the genus *Bacillus*. *Microbiology*, 134(7): 1847-1882.
- Sulaiman IM, Hsieh YH, Jacobs E, Miranda N, Simpson S, Kerdahi K (2018) Identification of *Lysinibacillus fusiformis* Isolated from Cosmetic Samples Using MALDI-TOF MS and 16S rRNA Sequencing Methods. *Journal of AOAC International*, 101(6): 1757-1762
- Prabha MS, Divakar K, Priya JDA, Selvam GP, Balasubramanian N, Gautam P (2015) Statistical analysis of production of protease and esterase by a newly isolated *Lysinibacillus fusiformis* AU01: purification and application of protease in

- sub-culturing cell lines. *Annals of microbiology*, 65(1): 33-46.
17. Coorevits A, Dinsdale AE, Heyrman J, Schumann P, Van Landschoot A, Logan NA, De Vos P (2012) *Lysinibacillus macroides* sp. nov., nom. rev. *International journal of systematic and evolutionary microbiology*, 62(5): 1121-1127.
 18. Delgado S, Rachid CT, Fernández E, Rychlik T, Alegría Á, Peixoto RS, Mayo B (2013) Diversity of thermophilic bacteria in raw, pasteurized and selectively-cultured milk, as assessed by culturing, PCR-DGGE and pyrosequencing. *Food microbiology*, 36(1): 103-111.
 19. Latorre I, Hwang S, Montalvo-Rodriguez R (2012) Isolation and molecular identification of landfill bacteria capable of growing on di-(2-ethylhexyl) phthalate and deteriorating PVC materials. *Journal of Environmental Science and Health, Part A*, 47(14): 2254-2262.
 20. Gopal N, Hill C, Ross PR, Beresford TP, Fenelon MA, Cotter PD (2015) The prevalence and control of *Bacillus* and related spore-forming bacteria in the dairy industry. *Frontiers in microbiology*, 6: 14-18.
 21. Sulaiman I M, Banerjee P, Hsieh YH, Miranda N, Simpson S, Kerdahi K (2018) Rapid Detection of *Staphylococcus aureus* and Related Species Isolated from Food, Environment, Cosmetics, a Medical Device, and Clinical Samples Using the VITEK MS Microbial Identification System. *Journal of AOAC International*, 101(4): 1135-1143.
 22. Hsieh YH, Wang YF, Moura H, Miranda N, Simpson S, Gowrishankar R, Sulaiman IM (2018) Application of Maldi-Tof MS Systems in the Rapid Identification of *Campylobacter* spp. of Public Health Importance. *Journal of AOAC International*, 101(3): 761-768.