



Molecular Detection of Some Virulence Factors Genes of *Proteus Mirabilis* That Isolated From Urinary Tract Infection

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Abstract

A total of (160) clinical specimen of urine collected from patients suffering from urinary tract infections at different ages and from both sex in different hospitals in Baghdad during the period from September 2017 to February 2018. were cultured on selective media MacConkey agar. Bacterial isolates were identified by a microscopic examination and diagnosed by using biochemical tests. A further identification by vitak 2system. The results showed that (103) samples were positive for bacteriological culture (64.6%), the percentage of *Proteus* spp were (54) urine sample (52.4%) and percentage of *Proteus mirabilis* isolates were (40) isolates (74%) while *Proteus vulgaris* were (14) isolates (25.9%) while the results indicated the isolation rate of *Proteus mirabilis* from females was (67%) which was higher than that of males (33%). Selection ten isolates of *P. mirabilis* which show highest urease activity and adhesion to uroepithelia cell. Adherence capability of the isolates was also tested in accordance to epithelial cells isolated from female urine all isolates were capable of adhering to epithelial cells. The genetic study included an extraction of genomic DNA from (10) *p. mirabilis*, then detection of virulence genes *ure C* that responsible for urease, *mrpA* for fimbrial and ability to adherence to uroepithelial cell for these isolated by using conventional PCR all isolated give positive result. Selection five from *ure c* and *mrp A* gene amplicons of *P. mirabilis* sent to Korea for direct sequencing. The nucleotide sequences of *ure c*, and *mrp A* genes were compared with the sequences of the *mrp A* and *ure c* gene using BLAST option of NCBI (Basic Local Alignment Search Tool, BLAST v. 2.2.15, www.ncbi.nlm.nih.gov). The result was 100% matching with the registrar globally.

Introduction

The genus *Proteus*, which was described for the first time by Hauser in 1885, belongs to the *Enterobacteriaceae* family. In this family it is placed in the tribe *Proteeae*, together with the genera *Morganella* and *Providencia* [1]. *Proteus* spp. consist of gram-negative, motile, Aerobic rod-shaped bacilli generally range from 0.3 to 1.0 μm in width and 0.6 to 6.0 μm in length [2]. The genus *Proteus* currently consists of five species which are *P. mirabilis*, *P. vulgaris*, *P. penneri*, *P. hauseri* and *P. myxofaciens* [3]. *P. mirabilis* expresses several virulence factor involved in infection like adhesions, flagella, toxins, quorum-sensing, enzymes and immune invasion [4]. *P. mirabilis*, once attached to urinary tract it infects the kidney more commonly than *E. coli*. It is best known for its ability to form stones in the bladder and kidney, as well as its ability to form crystalline biofilms on the outer surface and in the lumen of indwelling urinary

catheters [5].

Materials and Methods

Patients and Specimens

160 urine samples were collected from patients suffering from urinary tract infections: these samples taken from Medicine city/Baghdad teaching hospital, and Central Child hospital for the period from September 2017 to February 2018.

Bacterial Diagnosis

Isolation of *P. mirabilis* bacteria was performed by a surface streak procedure on both blood and MacConkey agar using calibrated loops and incubated aerobically at 37°C for 24 hours. Bacterial identification was made using biochemical test, namely indole, citrate, oxidase, catalase, urea hydrolysis, H₂S production, lactose fermentation

Detection of Urease Enzyme [6]

An amount of 10 µl sample solution was mixed with 990 µl urea determination solution in a small test tube, and the pH of the mixture was adjusted to 5.1 with 0.1 M HCl. The reaction was started by adding 2 µl *proteus mirabilis* suspension (about 2 × 10⁵ cells). After 20 min of incubation at room temperature, the color of the reaction solution was read at 588 nm.

Detection of Adhesion [7]

Preparation of the Selected Isolates of *P. mirabilis*

The cells of *Proteus* centrifuged, washed the pellet twice with PBS and the cells adjusted to contain about O.D 600 about 0.6 giving 1 × 10⁹ cells/ml by using viable count.

Preparation of Epithelial Cells

Uroepithelial cells were collected from urine of healthy females by centrifugation, then cells were washed three times with PBS before resuspension in PBS [8].

In Vitro Adhesion Test

Aliquot of bacterial suspension, and epithelial cells suspension, were incubated in shaker-incubator. Unattached bacterium were removed by centrifugation 3 times in PBS. The final pellet was resuspended during a drop of PBS, dropped onto a glass slide microscope and air dried. The glass slide was fixed with methanol : acetic acid (3:1) and stained with crystal violet .The number of attached bacteria was counted by light microscope.

Genomic DNA Extraction

DNA was extracted from ten isolates of *P. mirabilis* by using a commercial purification kit (Presto Mini Genomic DNA Kit, (Gene aid, Thailand).

F. PCR Amplifications

Detection of virulence genes was performed by amplifying the genes via PCR. Descriptions of the PCR primers used in this study are displayed in Table 1.G- Sequence analysis of *mrpA* and *ureC* genes for genotyping of isolates.

Table 1: Primers used in this study

Gene	Primer sequence (5'- 3')	Product size(bp)	Reference
<i>Ure C</i>	F: GTTATTCGTGATGGATGGG R : ATAAGGTGGTTACGCCAGA	317bp	9
<i>MrpA</i>	F:GAGCCATTCAATTAGGAATCCA R:AGCTCTGTACTIONCCTTGTACAGA	648bp	10

F*: Forward R**: Reverse

Table2: PCR program for amplification of *ure C* gene

No.	Stages	Temperature & time	
1	Initial denaturation	95 °C	2min
2	Denaturation	95 °C	30 sec
3	Annealing	56.2 °C	30 sec
4	Extension	72°C	1 min
5	Final extension	72°C	5 min

Table3: PCR program for amplification *mrpA* gene

No.	Stages	Temperature & time	
1	Initial denaturation	95°C	5min
2	Denaturation	95°C	1 min
3	Annealing	58°C	1 min
4	Extension	72°C	1 min
5	Final extension	72°C	7min

Results and Discussion

Isolation and Identification of *Proteus Mirabilis*

Loopful amount from these samples were inoculated on MacConkey agar and blood agar

and then incubated overnight at 37°C. A distinguishable swarming which is a unique characteristic for genus proteus was observed, which is considered as confirmatory phenomenon for genus proteus as previously described by [11].

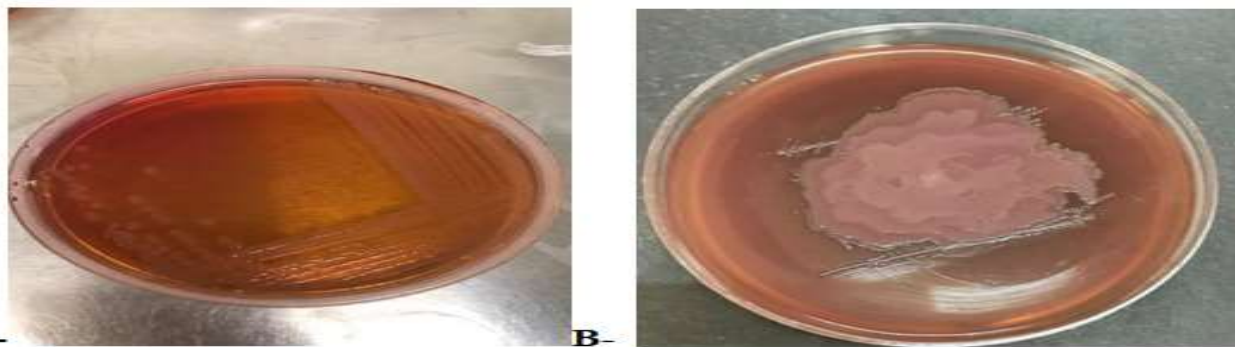


Figure1 A: *Proteus* colonies are non-lactose ferment or on MaCconkey agar after incubation at 37° C for 24 hrs. B: swarming phenomena on blood agar base

It was found that 103(64.3%) out of the total 160 samples collected gave positive results on MacConkey agar and Blood agar. These results were agreed with those reported by [12] who found that the percentage of positive cultures of urine samples were 64.6%. But such results were disagreed with those of [13] when found that percentage of positive culture of urine samples was (28.9%). The reason of the differences in percentage may be owed to differences in either size of samples or hospital locations as well as to the season and medications before sampling.

Result showed that 54 isolates belong to genus *Proteus* spp from the 103 positive cultures and, so the isolation percentage of *Proteus* from other bacteria of the UTI cases was (52.4%). This result was agreed with that of [14] who found that *Proteus* isolates were representing (60%) of the UTI cases tested.

But this result disagreed with [15] who found that isolation percentage of *Proteus* occurrence (17.8%). From the previous results, *P. mirabilis* represent 74% (40 isolates) while *P. vulgaris* appeared only in 25.9% (14 isolates). The present results with previous study done in by [16]. Who mentioned that *P. mirabilis* performed 77% while *p. vulgaris* performed 23%.

Determination of Urease activity

It has been noted that all 40 isolates under study were capable of producing urease. However, ten isolates of *Proteus mirabilis* have ability to produce urease as shown in Table (4). Maximum specific activity of enzyme was found to be 3.7 units/ml and the lowest specific activity of was found to be 0.9 units/ml. These results that obtained by [17] who detected highest specific activity of urease as 3.9 unit/ml.

Table 4: Urease activity produce by isolate of *P.mirabilis*

Isolates NO.	Absorbance at 588 nm	Ammonia conc. (u mole)	Specific activity (unit /ml)
1	0.381	106	3.5
2	0.176	49	1.6
3	0.384	107	3.5
4	0.273	76	2.5
5	0.186	52	1.7
6	0.372	104	3.4
7	0.224	62	2.0
8	0.357	99	3.3
9	0.121	34	1.1
10	0.362	100	3.3
11	0.375	104	3.4
12	0.210	59	1.9
13	0.281	78	2.6
14	0.216	60	2.0
15	0.134	37	1.2
16	0.382	106	3.5
17	0.250	70	2.3
18	0.399	111	3.7
19	0.375	104	3.4
20	0.284	79	2.6
21	0.140	39	1.3
22	0.218	61	2.0
23	0.281	78	2.6
24	0.115	32	1.0
25	0.350	97	3.2
26	0.102	29	0.9
27	0.112	31	1.0

28	0.213	59	1.9
29	0.236	66	2.2
30	0.221	62	2.0
31	0.211	59	1.9
32	0.218	61	2.0
33	0.152	42	1.4
34	0.169	47	1.5
35	0.371	103	3.4
36	0.150	42	1.4
37	0.098	27	0.9
38	0.110	31	1.0
39	0.315	88	2.9
40	0.099	28	0.9

Detection of Adhesion in *P. mirabilis*

Adhesive ability of *p.mirabilis* to epithelial cells isolated from female urine was compared by the frequency of distribution of bacteria on epithelial cells and by the mean number of bacteria adhering to (20) epithelial cells and considered as criteria for adhesive capability of cells, as long as adhering bacterial cells visible under light microscope and easy to count, all the isolates displayed differences in their adherence to human epithelial cells and showed a mean number of adhering bacteria ranging from(10.75 ± 0.63) to (18.00 ± 1.05)

bacteria /epithelial cell as indicated in Table(5) It was noticed that isolates no. 1, 3, 6, 8 , 10, 11, 16 , 18 , 19 and 35 displayed high adhesive capability with mean (18.00 ± 1.05), (16.50 ± 0.89), (17.00 ± 1.26), (15.75 ± 0.86), (17.50 ±1.16), (17.25 ± 0.97), (17.75 ± 1.38), (16.00 ± 1.07) and (16.50 ± 1.15) bacteria /epithelial cell respectively, while the isolate 24 displayed a lowest adhesive capability with mean (10.75 ± 0.63) bacteria /epithelial cell. These result [18].Who found that forty bacteria/ uroepithelial cell of adhesion averagedof *P. mirabilis* on uroepithelial cells.

Table 5: Adhesion average of *P. mirabilis* to uroepithelial cell

No. of <i>P.mirabilis</i> isolates	No. of adherent <i>P. mirabilis</i> to No. of epithelial cells				Mean No. of adherent <i>P. mirabilis</i> /cells ± SD
	0	1-5	6-20	>20	
1	0	1	6	13	18.00 ± 1.05
2	2	10	3	5	12.75 ± 0.63
3	0	4	6	10	16.50 ± 0.89
4	0	6	14	0	13.50 ± 0.52
5	1	12	5	2	12.00 ± 0.48
6	0	2	8	10	17.00 ± 1.26
7	2	8	7	3	12.75 ± 0.73
8	2	3	5	10	15.75 ± 0.86
9	2	13	5	0	10.75 ± 0.69
10	0	2	6	12	17.50 ± 1.16
11	0	2	7	11	17.25 ± 0.97
12	0	6	10	4	14.50 ± 0.61
13	0	12	6	2	12.50 ± 0.74
14	2	9	4	5	13.00 ± 0.68
15	2	9	6	3	12.50 ± 0.71
16	0	2	5	13	17.75 ± 1.38
17	0	7	13	0	13.25 ± 0.74
18	2	2	6	10	16.00 ± 1.07
19	2	3	5	10	15.75 ± 0.86
20	0	8	8	4	14.00 ± 0.82
21	1	9	6	4	13.25 ± 0.77
22	1	8	7	4	13.50 ± 0.82
23	0	10	7	3	13.25 ± 0.75

24	2	13	5	0	10.75 ± 0.63
25	0	6	14	0	13.50 ± 0.78
26	1	12	5	2	12.00 ± 0.61
27	0	6	10	4	14.50 ± 0.95
28	0	12	5	3	12.75 ± 0.72
29	3	10	7	0	11.00 ± 0.47
30	1	12	5	2	12.00 ± 0.51
31	1	6	8	5	14.25 ± 0.82
32	1	8	6	5	13.75 ± 0.76
33	2	6	7	5	13.75 ± 0.81
34	0	8	7	5	14.25 ± 0.79
35	0	4	6	10	16.50 ± 1.15
36	1	12	5	2	12.00 ± 0.49
37	1	10	6	3	12.75 ± 0.77
38	2	12	6	0	11.00 ± 0.46
39	0	12	6	2	12.50 ± 0.65
040	0	6	9	5	14.75 0.92

Amplification of ure C by Conventional PCR Techniques

Amplification of urease gene was done using specific PCR primer. Results shown in figure indicate successful amplification of the gene

for all isolates as indicated by the presence of band with molecular weight 317 bp. Regarding Ure C gene, these results are with a previous study by[19] who indicating the presence of Ure C about 96.6% isolates obtained from UTI patients.

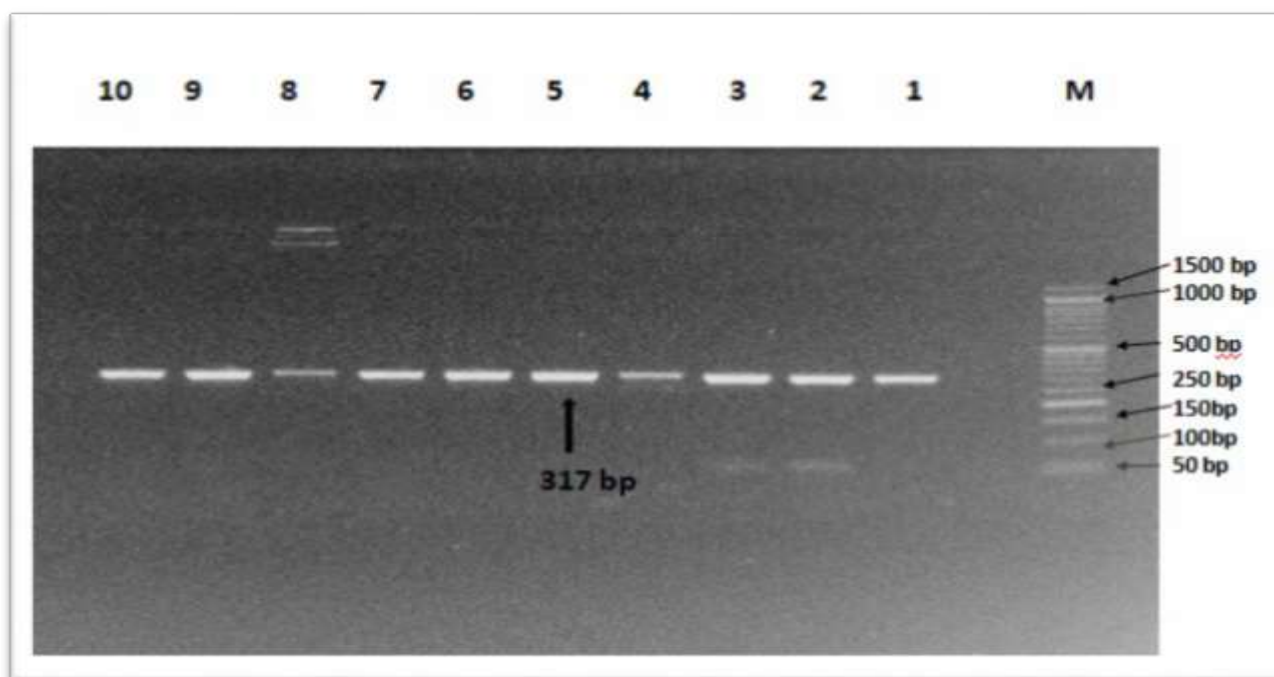


Figure 2: Gel electrophoresis for amplified ure C gene of proteus mirabilis Lane M ladder (100 bp), Line: 1-10 represent isolates 1, 3, 6, 8, 10, 11, 16, 18, 19 and 35 respectively

Amplification of mrp A by Conventional PCR Techniques

Amplification of mrpA gene was done using specific PCR primer. Results shown in Figure (3) indicate successful amplification of the gene for all isolates as indicated by the

presence of band with molecular weight 648bp. This results [20] who found that *P. mirabilis mrpA* fimbrial gene was present in all isolates. This particular type of fimbriae is frequently related to *P. mirabilis* UTI pathogenesis.

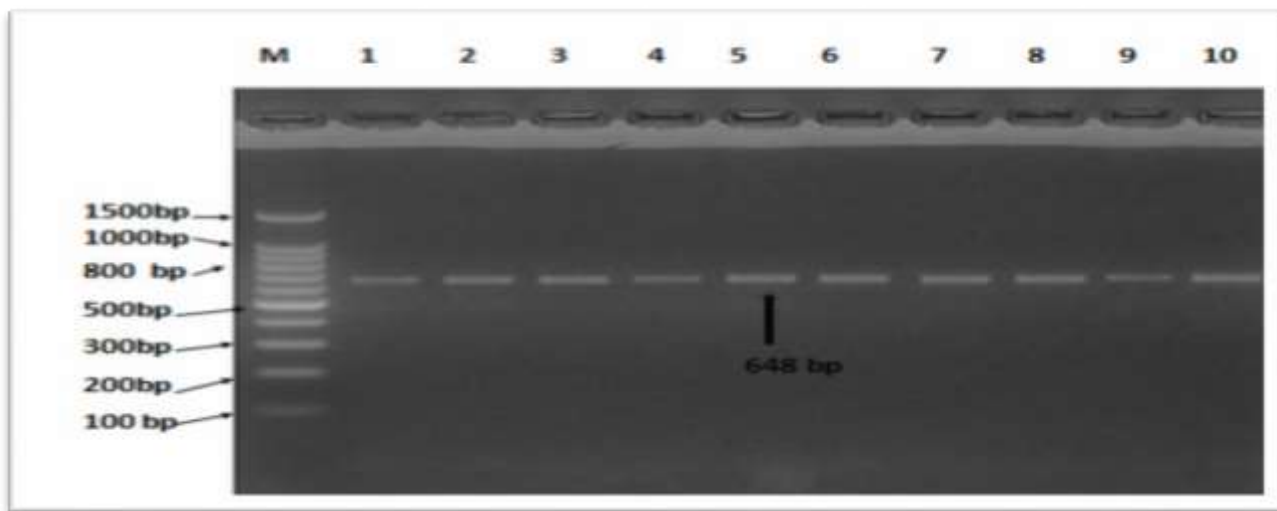


Figure 3: Gel electrophoresis for amplified *mrpA* gene of proteus mirabilis Lane M ladder (100 bp), Line: 1-10 represent isolates 1, 3, 6, 8, 10, 11, 16, 18, 19 and 35 respectively

Sequence Analysis of *mrpA* gene

The sequence homology of *mrpA* gene was obtained by comparison with closest blast sequence in gene bank. Similarity was

calculated using FASTA method. Data shown in table (6) indicate as much as 100% similarity of four isolates with that sequence of *P.mirabilis* AR-0029. however, isolate 19 shows 100% homology with that of *P.mirabilis* AR-0156.

Table 6: Average similarity percentage of *mrpA* sequences of *P. mirabilis* isolated from UTI patients in comparison to sequence obtained from NCBI blast

No. of local isolate	NCBI Blast Alignment	<i>mrpA</i> Sequence Identity (%)
1	<i>P.mirabilis</i> AR-0029	100%
2	<i>P.mirabilis</i> AR-0029	100%
3	<i>P.mirabilis</i> AR-0029	100%
4	<i>P.mirabilis</i> AR-0029	100%
5	<i>P.mirabilis</i> AR-0156	100%

These results were in agreement with several other investigations namely; Pathirana, (2018) show that *mrpA* gene of *P. mirabilis* represents 96.40% of mean similarity and pet turtles to human respiratory isolates using the same gene of *P. mirabilis* represents 94.90% of mean similarity also [20] found similarity in *mrpA* amplicon sequences of chicken to human isolates.

The sequence homology of ure c gene was obtained by comparison with closest blast sequence in gene bank. Similarity was calculated using FASTA method. Data shown in Table (7) indicate as much as 100% similarity of two isolated with that sequence of p.mirabilisAR-0159. However, two isolate show 100% homology with that of *P. mirabilis* AR-K1609 and other isolate show 100% homology with that of *P.mirabilis*AR-379

Sequence Analysis of *ure C* Gene

Table 7: Average similarity percentage of *ure C* sequence of *P.mirabilis* isolated from UTI patients in comparison to sequence obtained from NCBI blast

No. of Local Isolate	NCBI Blast Alignment	<i>Ure C</i> Sequence Identity (%)
1	<i>P.mirabilis</i> AR-0159	100%
2	<i>P.mirabilis</i> AR-379	100%
3	<i>P.mirabilis</i> AR-0159	100%
4	<i>P.mirabilis</i> AR-k1609	100%
5	<i>P.mirabilis</i> AR-K1609	100%

These results were several other investigation who [21] show *ure c* gene of *P. mirabilis*

represents of obtained were sequenced and BLAST search, which revealed 100% identity.

References

- Rozalski A, Staczek P (2011) Proteus. In: Molecular detection of human bacterial pathogens. In: D. Liu (ed.), CRC Press, Taylor and Francis Group. Boca Raton, 981-996.
- Abbott SL (2007) Klebsiella, Enterobacter, Citrobacter, Serratia, Plesiomonas, and other Enterobacteriaceae. In P. R. Murray, E. J. Baron, J.H. Jorgensen, M. L. Landry & M. A.

- Pfaller (Eds.), Manual of Clinical Microbiology (9th ed., pp. 698-711). Washington, USA: ASM Press.
3. O'hara, MC, Brenner WF, Miller MJ (2000) Classification, identification, and clinical significance of *Proteus*, *Providencia*, and *Morganella*. *Clinical Microbiology Reviews*, 13: 534-546.
 4. Baldo C, Rocha SP (2014) Virulence factors of uropathogenic *Proteus mirabilis*. *International journal of Scientific and technology research*, 3(11):24-27.
 5. Pearson M, Sebahia M, Churcher C, Quail MA, Seshasayee AS, Luscombe N, Abdellah Z, Arrosmith C, Atkin B, Chilling worth TH, Hauser K, Jagels S, Moule K, Mungall H, Norbertczak E, Rabinowitsch D, Walker S, Whithead NR, Thomson PN, Rather J, Parkhill Mobley HL (2008) Complete genome sequence of uropathogenic *Proteus mirabilis*, a master of both adherence and motility, *Journal of Bacteriology*, 190: 4027-4037.
 6. Lin YL, Chen CT, Lin SC, Lee C, Kuo HS, Shih CM, Chan EC (2000) A simple method to determine urea concentration using intact *Helicobacter pylori* and BromoCresol Purple as a pH indicator. *Biotechnology Letters*, 22(13): 1077-1079.
 7. Iwahi T, Abe Y, Tsuchiya K (1982) Virulence of *E. coli* in a sending urinary tract infection in mice. *Journal Medical Microbiology*, 15: 303-316.
 8. Chan RC, Burce AW, Raid G (1984) Adherence of cervical, vaginal and distal urethral normal microbial flora to human uroepithelial cells and inhibition of adherence of gram-negative uropathogens by competitive exclusion. *The Journal of Urology*, 131:596-60.
 9. Stankowska D, Kwinkowski M, Kaca W (2008) Quantification of *Proteus mirabilis* virulence factors and modulation by acylated homoserine lactones. *Journal Microbiol Immunol Infectious*, 41(3): 243-253.
 10. Sergio PD, Rocha P, Aurora M, Marcio A, Julia M, Roxane MF, Michele RL, Cristiano G, Jacinta S (2007) Aggregative adherence of uropathogenic *Proteus mirabilis* to cultured epithelial cells. *FEMS Immunol Medical Microbiol.*, 51: 319-3.
 11. Liaw SJ, La HC, Ho SW, Wang WB (2000) Inhibition of virulence factor expression and swarming differentiation in *Proteus mirabilis* by *P. nitrophenylglycerol*. *Journal of Medical Microbiology*, 49:725-731.
 12. Al-Aabideen R (2005) Effect of Probiotic on Motility Factors and Swarming Phenomenon of *Proteus mirabilis*. M.Sc. Thesis. Collage of science. Al-Nahrain University.
 13. Al-Kabby A (2007) Extraction and Partial Purification for Fimbriae from *Proteus mirabilis* and Study Their Role in Adhesion to Uroepithelial Cells. M.Sc. degree. College of Sciences, Baghdad University.
 14. Ahmed DA (2015). Prevalence of *Proteus* spp. in some hospitals in Baghdad City. *Iraqi Journal of Science*, 56(1C): 665-672.
 15. Adnan PM, Aziz PI H Al-Deresawi P MS (2014) Molecular detection of *Proteus mirabilis* using PCR technique among urinary tract infection patients. *Iraqi Journal of Biotechnology*, 13(2): 35-47.
 16. Yassen LT, Khelkal IN (2015) Effect of some fatty acid on virulence factors of *proteus mirabilis*. *Internal journal of advance biological research*, 5(2): 108-117.
 17. Awadh A (2004) Extraction and characterization of urease enzyme of *Proteus* spp. isolated from patients with urinary tract infection in Yemen. Ph.D. Thesis, Collage of Science, Al-Mustansirya University.
 18. Al-Bassam WW (2012) Genetic Study on Swarming Phenomenon and Adherence capacity of *Proteus mirabilis*. *Iraqi Journal of Biotechnology*, 11(2): 537-549.
 19. Pathirana HN, De Silva, BC Wimalasena, SH Hossain, S Heo GJ (2018) Comparison of virulence genes in *Proteus* species isolated from human and pet turtle. *Iranian journal of veterinary research*, 19(1): 48.
 20. Barbour EK, Hajj ZG, Hamadeh S, Shaib HA, Farran MT, Araj G, Kumosani T (2012) Comparison of phenotypic and virulence genes characteristics in human and chicken isolates of *Proteus mirabilis*. *Pathogens and global health*, 106(6): 352-357.
 21. Pattanayak S, Kumar P, R Sahoo, MK Paul A Sahoo PK (2018) First field-based evidence of association of *Proteus mirabilis* causing large scale mortality in Indian major carp farming. *Aquaculture*, 495: 435-442.